551-1119-00L Microbial Community Genomics

16S rRNA amplicon data analysis using DADA2

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The 16S rRNA gene

30S small subunit

12

Part of prokaryotic ribosomes

50S large subunit (33 proteins)	<u>5S</u> : 120 nt <u>23S</u> : 2906 nt
30S small subunit (22 proteins)	<u>16S</u> : 1542 nt

16S rRNA present in all prokaryotes

blue: ribosomal proteins gold: 16S rRNA

V6

V8

conserved regions and variable regions

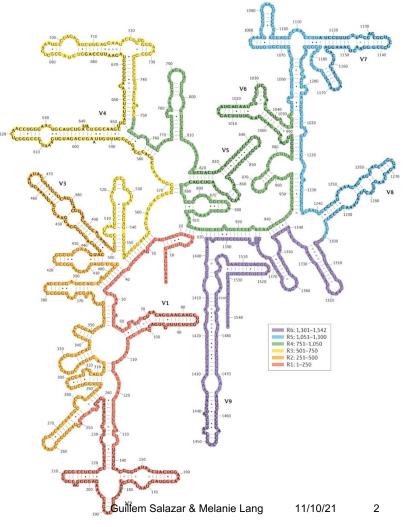
V3

0 100 200 300 400 500 600 700 800 900 1000 1100 1200 1300 1400 1500 bp

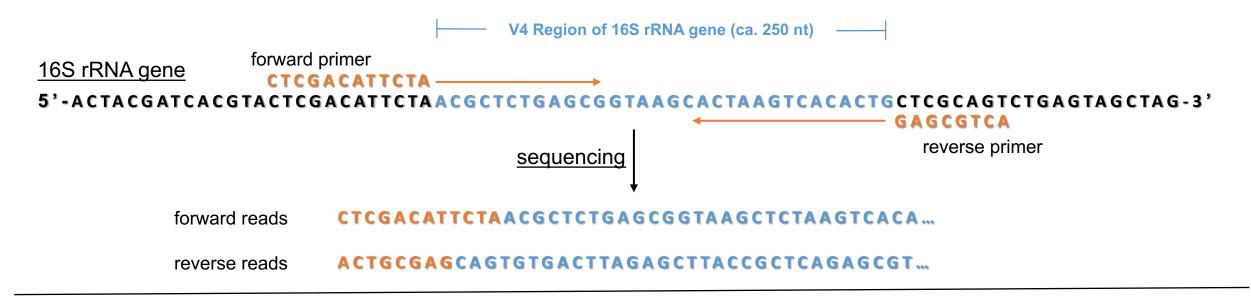
/5

V4



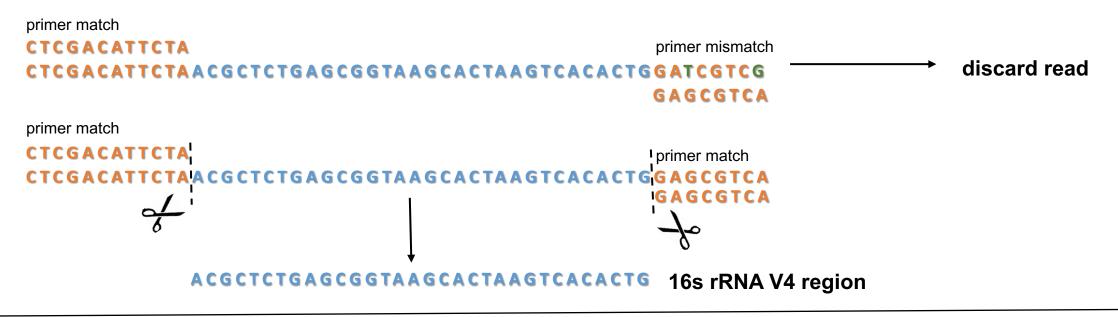


Generation of 16S rRNA gene PCR amplicons



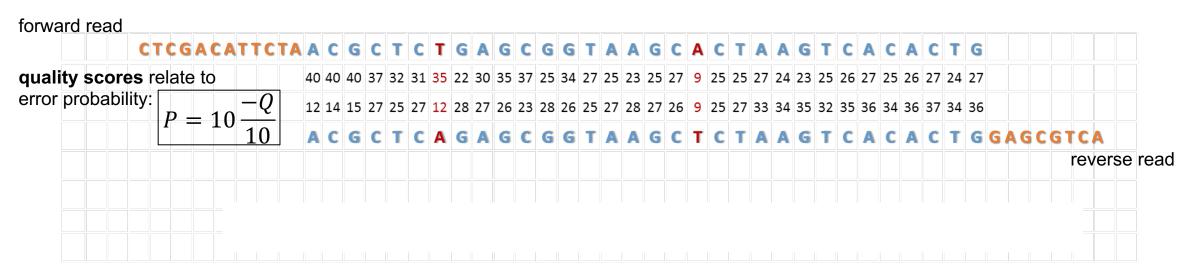
- PCR products (amplicons) are used for sequencing
- The sequencing output are forward reads and reverse reads
- Sequencing reads are saved as plain (or compressed) text files in the fastq format
 - see: https://en.wikipedia.org/wiki/FASTQ_format

Quality control reads: primer match filtering



- Forward and reverse primer sequences are aligned to the read.
- If both primers perfectly match, the read is used for further steps, otherwise the read is discarded.
- This assures that all reads start at the same 16S position, which is mandatory for the pipeline to work.

Quality scores of amplicon reads



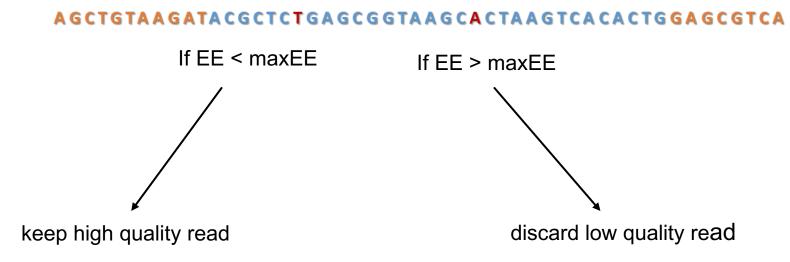
- Each base of the forward and reverse have a quality (Q) score:
 - the higher the Q score the higher the probability that the base call is correct

Quality control of amplicon reads: error filtering

• Quality filtering by maximum expected errors (maxEE) should be performed as a first processing step

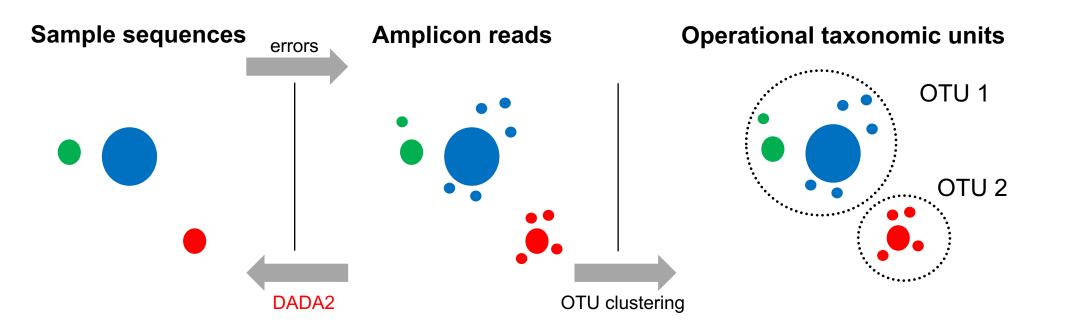
EE = expected errors = sum of error probability (sum of P)

- small EE = high quality; large EE = low quality
- By setting a maximum expected errors (maxEE) threshold, we can discard reads with EE > maxEE



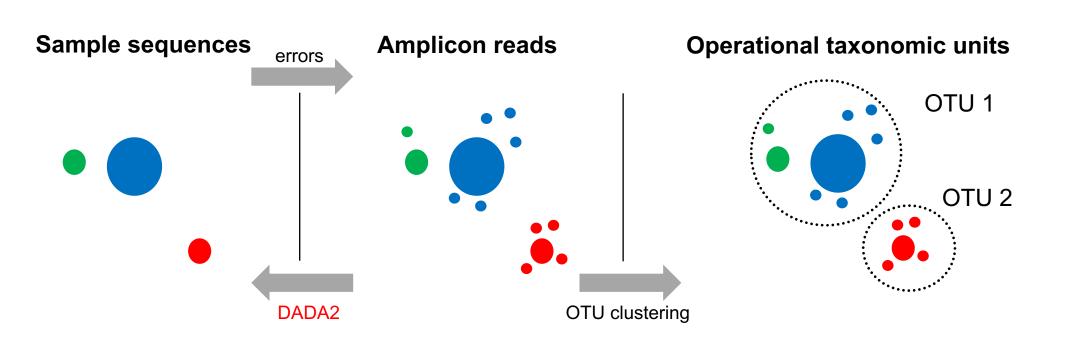
The Divisive Amplicon Denoising Algorithm (DADA)

- The core denoising algorithm in the DADA2 R package is built on a model of the errors in Illumina-sequenced amplicon reads.
- This error model quantifies the rate λ_{ji} at which an amplicon read with sequence *i* is produced from sample sequence *j* as a function of sequence composition and quality.



The Divisive Amplicon Denoising Algorithm (DADA)

 DADA2 tries to decipher whether each amplicon is a true biological sequence or is the result of an error because of the sequencing process

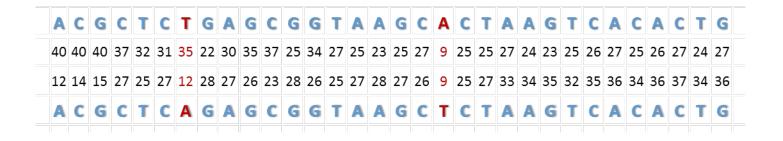


DADA2: the error model

The rate at which an amplicon read with sequence *i* is produced from sample sequence *j* is reduced to the product over the transition probabilities between the *L* aligned nucleotides.

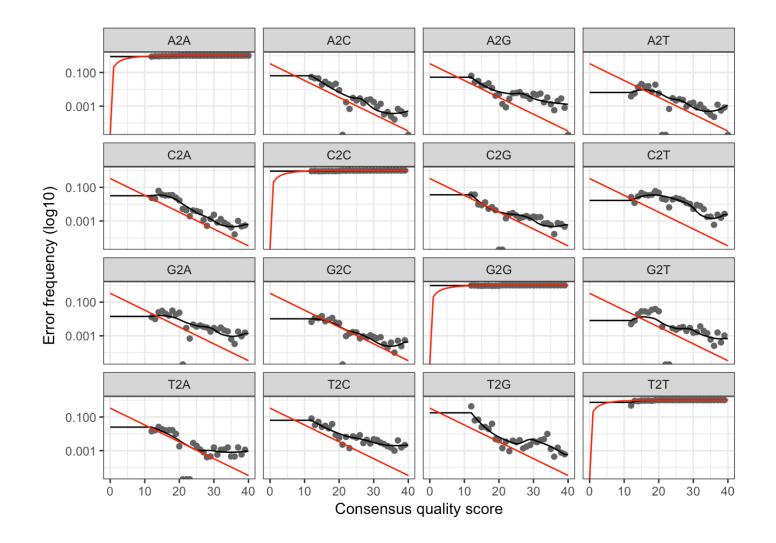
$$\lambda_{ji} = \prod_{l=0}^{L} p(j(l) \rightarrow i(l), q_i(l))$$

 The transition probability between aligned nucleotides is allowed to depend on the original nucleotide, substituting nucleotide, and associated quality score, for example, p(A→T, 9)



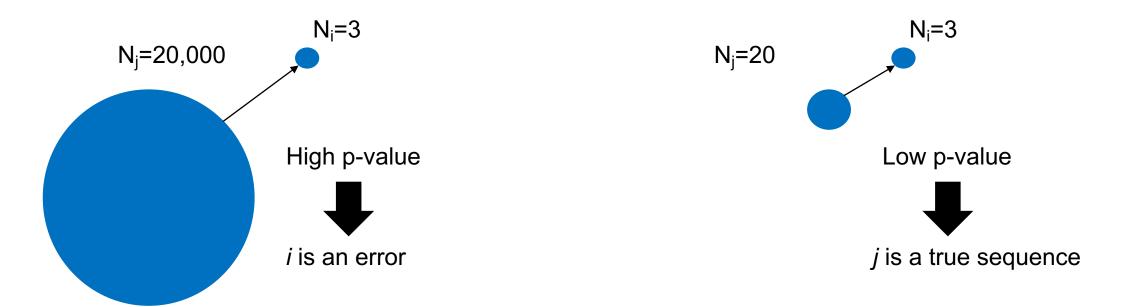
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DADA2: the error model



DADA2: the abundance p-value

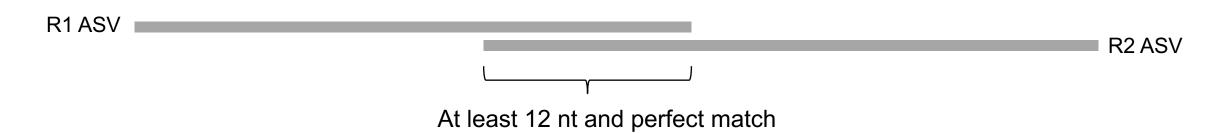
- The abundance *p*-value quantifies the notion that sequence *i* is too abundant to be explained by errors in amplicon sequencing.
- It measures the probability of a given amplicon abundance given the model. That is, the likelihood that an amplicon is produced n times because errors.



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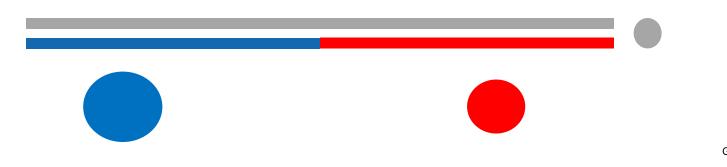
Merging and chimera removal

Merging:

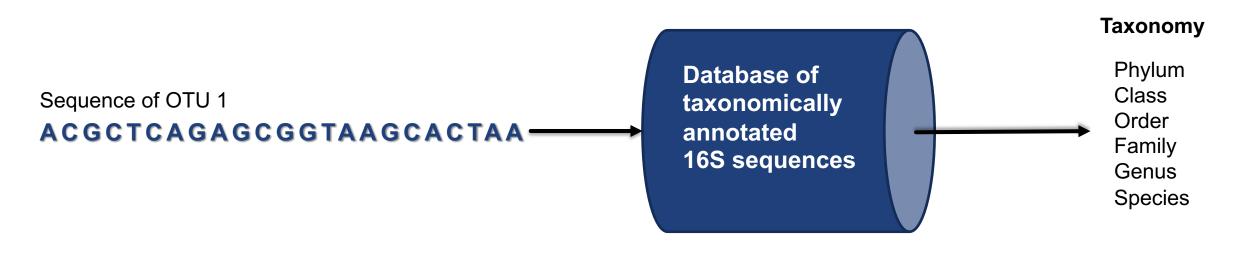


• Chimera detection and removal:

Chimeric sequences are identified if they can be exactly reconstructed by combining a left-segment and a rightsegment from two more abundant "parent" sequences



Taxonomic annotation of ASVs



- Prediction of ASV taxonomy
- Each is compared to a database of annotated 16S rRNA gene sequences
- Sequences are classified to a phylum, class, family etc.