551-1119-00L Microbial Community Genomics

Introduction to Comparative Genomics

DBIOL

Sequencing technologies: an historical perspective

- 1953: Discovery of the structure of DNA
- o 1965: "Sequencing" of the first tRNA
- o 1972: Sequencing of first complete gene (coat protein of bacteriophage MS2)
- o 1977: Release of "**chain termination method**" à **FIRST GENERATION SEQUENCING**
- 1996: Beginning of **SECOND or NEXT-GENERATION SEQUENCING**
- o 2005: Implementation of pyrosequencing in automated system
- o 2007: Illumina acquires Solexa
- 2010: Beginning of **THIRD-GENERATION SEQUENCING**

o 1953: Discovery of the structure of DNA

Francis Crick

Rosalind Franklin

James Watson

Maurice Wilkins

- o 1953: Discovery of the structure of DNA
- o 1965: "Sequencing" of the first tRNA

 \rightarrow use of ribonucleases with cleaving sites at specific nucleotides

 \rightarrow reconstruction of the original nucleotide sequence by determining the order in which small fragments occurred in the tRNA molecule

| |

Robert W. Holley

- o 1953: Discovery of the structure of DNA
- o 1965: "Sequencing" of the first tRNA

 \rightarrow use of ribonucleases with cleaving sites at specific nucleotides

 \rightarrow reconstruction of the original nucleotide sequence by determining the order in which small fragments occurred in the tRNA molecule

o 1972: Sequencing of first complete gene (coat protein of bacteriophage MS2) via RNAse digestion and isolation of oligonucleotides

| |

Robert W. Holley

Walter Fiers

- o 1953: Discovery of the structure of DNA
- o 1965: "Sequencing" of the first tRNA

 \rightarrow use of ribonucleases with cleaving sites at specific nucleotides

 \rightarrow reconstruction of the original nucleotide sequence by determining the order in which small fragments occurred in the tRNA molecule

- o 1972: Sequencing of first complete gene (coat protein of bacteriophage MS2) via RNAse digestion and isolation of oligonucleotides
- o 1977: Release of "**chain termination method**" utilizing radiolabeled partially digested fragments \rightarrow **FIRST GENERATION SEQUENCING**

Robert W. Holley

Walter Fiers

Frederick Sanger

- o 1953: Discovery of the structure of DNA
- o 1965: "Sequencing" of the first tRNA

 \rightarrow use of ribonucleases with cleaving sites at specific nucleotides

 \rightarrow reconstruction of the original nucleotide sequence by determining the order in which small fragments occurred in the tRNA molecule

- o 1972: Sequencing of first complete gene (coat protein of bacteriophage MS2) via RNAse digestion and isolation of oligonucleotides
- o 1977: Release of "**chain termination method**" utilizing radiolabeled partially digested fragments \rightarrow **FIRST GENERATION SEQUENCING**

Robert W. Holley

Walter Fiers

 \rightarrow Main sequencing technology for next 25 years \rightarrow Key innovations mainly in automation of wetlab and data analysis pipelines

DBIOL

Frederick Sanger

- o 1996: Beginning of **NEXT-GENERATION SEQUENCING**
	- \rightarrow Pyrosequencing

nucleotides

DBIOL https://the-dna-universe.com/2020/11/02/a-journey-through-the-history-of-dna-sequencing/

DBIOL

- o 1996: Beginning of **NEXT-GENERATION SEQUENCING**
	- \rightarrow Pyrosequencing

| |

nucleotides

https://www.ebi.ac.uk/training/online/courses/functional-genomics-ii-common-technologies-and-dataanalysis-methods/next-generation-sequencing/454-sequencing/

- o 1996: Beginning of **NEXT-GENERATION SEQUENCING**
	- \rightarrow Pyrosequencing

nucleotides

DBIOL

- o 1996: Beginning of **NEXT-GENERATION SEQUENCING**
	- \rightarrow Pyrosequencing

nucleotides

DBIOL

- o 1996: Beginning of **NEXT-GENERATION SEQUENCING** \rightarrow Pyrosequencing
- o 2005: Implementation of pyrosequencing in automated system
	- \rightarrow 454 sequencing platform

Roche 454 Sequencing System

| |

- o 1996: Beginning of **NEXT-GENERATION SEQUENCING** \rightarrow Pyrosequencing
- o 2005: Implementation of pyrosequencing in automated system
	- \rightarrow 454 sequencing platform
- o 2007: Illumina acquires Solexa
	- \rightarrow Advanced sequencing technology
	- \rightarrow Improved throughput
		- \rightarrow In each cycle, one dNTP is incorporated into the reaction and it's fluorescent signal captured in an image
		- \rightarrow Process is repeated until a full "read" is assembled

Illumina MiSeq Sequencing platform

| |

AGTTCGA

glass flow cell

 \rightarrow Flow cell is flooded with fluorescently labelled dNTPs and polymerase

DBIOL

CTACGAT... \rightarrow Translation of thousands of fluorescent images into a sequence read

https://the-dna-universe.com/2020/11/02/a-journey-through-the-history-of-dna-sequencing/ https://www.ebi.ac.uk/training/online/courses/functional-genomics-ii-common-technologies-and-data-analysis-methods/next-generationsequencing/454-sequencing/

Improvements in DNA sequencing: Some numbers…

experience in production l *Moore's law is an observation and projection of a historical trend. Rather than a law of physics, it is an empirical relationship linked to gains from

o 2010: Beginning of **THIRD-GENERATION SEQUENCING**

 \rightarrow PacBio sequencing (Pacific Biosciences, Inc.)

| |

PacBio RSII sequencer

 \rightarrow polymerase immobilized at the bottom of a "well" (zero-mode waveguide ZMW) in a SMRTcell

 \rightarrow Incorporation of fluorescent dNTPs produces a base-specific light pulse

 \rightarrow Replication process in all ZMWs is recorded as a "movie" in real-time

- o 2010: Beginning of **THIRD-GENERATION SEQUENCING**
	- \rightarrow PacBio sequencing (Pacific Biosciences, Inc.)

| |

PacBio RSII sequencer

https://the-dna-universe.com/2020/11/02/a-journey-through-the-history-of-dna-sequencing/ Rhoads & Au (2015) PacBio sequencing and its applications *Genomics, Proteomics & Bioinformatics* 13(5): 278-289

DBIOL

o 2010: Beginning of **THIRD-GENERATION SEQUENCING**

 \rightarrow PacBio sequencing (Pacific Biosciences, Inc.)

 \rightarrow Nanopore sequencing (Oxford Nanopore Technologies)

| |

Nanopore MinION

- \rightarrow single-stranded DNA/RNA molecules pass through protein nanopore
- \rightarrow Each nucleotide that passes the pore leads to a different change in electrical current across pore
- \rightarrow Resulting signal is decoded to provide sequence information

https://the-dna-universe.com/2020/11/02/a-journey-through-the-history-of-dna-sequencing/

https://www.sciencedirect.com/topics/neuroscience/nanopore-sequencing

https://nanoporetech.com/applications/dna-nanopore-sequencing

DBIOL

o 2010: Beginning of **THIRD-GENERATION SEQUENCING**

 \rightarrow PacBio sequencing (Pacific Biosciences, Inc.)

 \rightarrow Nanopore sequencing (Oxford Nanopore Technologies)

| |

Nanopore MinION

- \rightarrow single-stranded DNA/RNA molecules pass through protein nanopore
- \rightarrow Each nucleotide that passes the pore leads to a different change in electrical current across pore
- \rightarrow Resulting signal is decoded to provide sequence information

https://the-dna-universe.com/2020/11/02/a-journey-through-the-history-of-dna-sequencing/

https://www.sciencedirect.com/topics/neuroscience/nanopore-sequencing

https://nanoporetech.com/applications/dna-nanopore-sequencing

Genome sequencing: an historical perspections

https://doi.org/10.1016/

See also: https://www.nature.com/immersive/d42859-020-00099-0/index.html for milestones of genome sequencing.

DBIOL

The great plate count anomaly

• The "**great plate count anomaly**" is the term we use to describe the observation that microscopic cell counts are significantly higher than corresponding counts of "*colony forming units*" on agar plates.

FIGURE 6.12. The great plate count anomaly. Plate counts of cells obtained by cultivation are usually much lower, sometimes by orders of magnitude, than those from direct cell counts under a microscope. Possible reasons are (1) the differing nutritional requirements of the organism, (2) the organism may enter a noncultivatable resting state, or (3) the organism may rely on other organisms and thus cannot be cultivated in isolation.

Evolution © 2007 Cold Spring Harbor Laboratory Press

Marine microbial diversity: an historical perspective

Salazar & Sunagawa, 2017, Current Biology

Genome reconstruction from metagenomes

DBIOL

Recent findings in marine microbial genomics

- o Discovery of hidden clades
	- o Asgard Archaea
	- o Candidate phyla radiation (CPR)
	- o DPANN
- o Discovery of new metabolisms: COMAMMOX
- o Re-definition of known metabolisms: the case of nitrogen fixation

^o **Discovery of hidden clades: Asgard archaea**

Asgard archaea or Asgardarchaeota is a proposed superphylum consisting of a group of archaea that includes Lokiarchaeota, Thorarchaeota, Odinarchaeota, and Heimdallarchaeota. It appears the eukaryotes have emerged within the Asgard, which supports the twodomain system of classification over the three-domain system.

Global distribution of metagenomic-assembled sequences of Asgard archaea. Asgard metagenomicassembled genomes from NCBI Assembly and MG-RAST databases were recorded for information related to location and environmental context of sampling (November 2018)

Discovery of hidden clades: Candidate phyla radiation

The **candidate phyla radiation (CPR group)** is a large evolutionary radiation of bacterial lineages whose members are mostly uncultivated and only known from metagenomics and single cell sequencing.

CPR lineages are generally characterized as:

- having **small genomes** and
- **lacking several biosynthetic pathways and ribosomal proteins**.

This has led to the speculation that they are likely obligate symbionts.

Discovery of hidden clades: DPANN

DPANN is a superphylum of Archaea first proposed in 2013. They are known as nanoarchaea or ultra-small archaea due to their smaller size. They exhibit limited metabolic capacities reflected in the fact that many lack central biosynthetic pathways for nucleotides, aminoacids, and lipids. They are mostly anaerobic and cannot be cultivated.

RESEARCH ARTICLE | MICROBIOLOGY | C

Insight into the symbiotic lifestyle of DPANN archaea revealed by cultivation and genome analyses

Hiroyuki D. Sakai \bullet , Naswandi Nur, Shingo Kato \bullet , +s, and Norio Kurosawa \bullet \circ Authors Info & Affiliations Edited by Edward DeLong, Daniel K. Inouye Center for Microbial Oceanography: Research and Education, University of Hawaii at Manoa, Honolulu, HI; received August 26, 2021; accepted November 12, 2021

January 12, 2022 | 119 (3) e2115449119 | https://doi.org/10.1073/pnas.2115449119

Significance

The DPANN superphylum is a grouping of symbiotic microorganisms categorized based on their genomic contents and a few examples of cultivation experiments. Although the genome information of DPANN archaea is increasing year by year, most of them have remained uncultivated, limiting our knowledge of these organisms. Herein, a thermoacidophilic symbiotic archaeon (ARM-1) from the DPANN superphylum was successfully cultivated and characterized. We determined its physiological, morphological, and genomic characteristics in detail and obtained experimental evidence of the symbiotic lifestyle of this archaeon. Notably, ARM-1 is a symbiotic archaeal strain that showed dependence on a range of host species in a laboratory culture. The results significantly contribute to the true understanding of the physiology and ecology of DPANN archaea.

Discovery of new metabolisms: COMAMMOX

Comammox (COMplete AMMonia OXidation) is the name attributed to an organism that can convert ammonia into nitrite and then into nitrate through the process of nitrification.

Nitrification has traditionally thought to be a two-step process, where ammonia-oxidizing bacteria and archaea oxidize ammonia to nitrite and then nitrite-oxidizing bacteria convert to nitrate.

Complete conversion of ammonia into nitrate by a single microorganism was first predicted in 2006. In 2015 the presence of microorganisms that could carry out both conversion processes was discovered within the genus Nitrospira, and the nitrogen cycle was updated.

Published: 26 November 2015

Complete nitrification by Nitrospira bacteria

Holger Daims, Elena V. Lebedeva, Petra Pjevac, Ping Han, Craig Herbold, Mads Albertsen, Nico Jehmlich, Marton Palatinszky, Julia Vierheilig, Alexandr Bulaev, Rasmus H. Kirkegaard, Martin von Bergen, Thomas Rattei, Bernd Bendinger, Per H. Nielsen & Michael Wagner

Nature 528, 504-509 (2015) Cite this article 47k Accesses | 1240 Citations | 186 Altmetric | Metrics Published: 26 November 2015

Complete nitrification by a single microorganism

Maartje A. H. J. van Kessel, Daan R. Speth, Mads Albertsen, Per H. Nielsen, Huub J. M. Op den Camp, Boran Kartal, Mike S. M. Jetten & Sebastian Lücker

Nature 528, 555-559 (2015) Cite this article 35k Accesses | 920 Citations | 124 Altmetric | Metrics

o **Re-definition of known metabolisms: the case of nitrogen fixation**

Nitrogen fixation is the conversion of molecular nitrogen into ammonia or related nitrogenous compounds, typically in soil or aquatic systems. Biological nitrogen fixation or diazotrophy is an important microbials mediated process that converts dinitrogen gas to ammonia using the nitrogenase protein complex.

Two very recent findings, both based on the reconstruction of marine MAGs, have re-defined our knowledge of diazotrophy:

- The existence and relevance of marine **Heterotrophic Bacterial Diazotrophs** (HBDs)
- The existence of **non-diazotrophic Trichodesmium** (a genus previously though to be strictly diazotrophic).

RESEARCH ARTICLE | MICROBIOLOGY | @

 $f \vee in \n\mathbb{Z}$

Discovery of nondiazotrophic Trichodesmium species abundant and widespread in the open ocean

Tom O. Delmont ¹ ⊠ Authors Info & Affiliations

Edited by Paul G. Falkowski, Rutgers, The State University of New Jersey, New Brunswick, NJ, and approved September 13, 2021 (received fo review July 8, 2021)

November 8, 2021 | 118 (46) e2112355118 https://doi.org/10.1073/pnas.2112355118

Article | Open Access | Published: 11 June 2018

Nitrogen-fixing populations of Planctomycetes and Proteobacteria are abundant in surface ocean metagenomes

Tom O. Delmont ⊠, Christopher Quince, Alon Shaiber, Özcan C. Esen, Sonny TM Lee, Michael S. Rappé, Sandra L. McLellan, Sebastian Lücker & A. Murat Eren ⊠

Nature Microbiology 3, 804-813 (2018) Cite this article