

# Assembly-free shotgun metagenomic profiling

15 November 2022

**Alessio Milanese**  
Postdoc in the  
Sunagawa group (ETH)

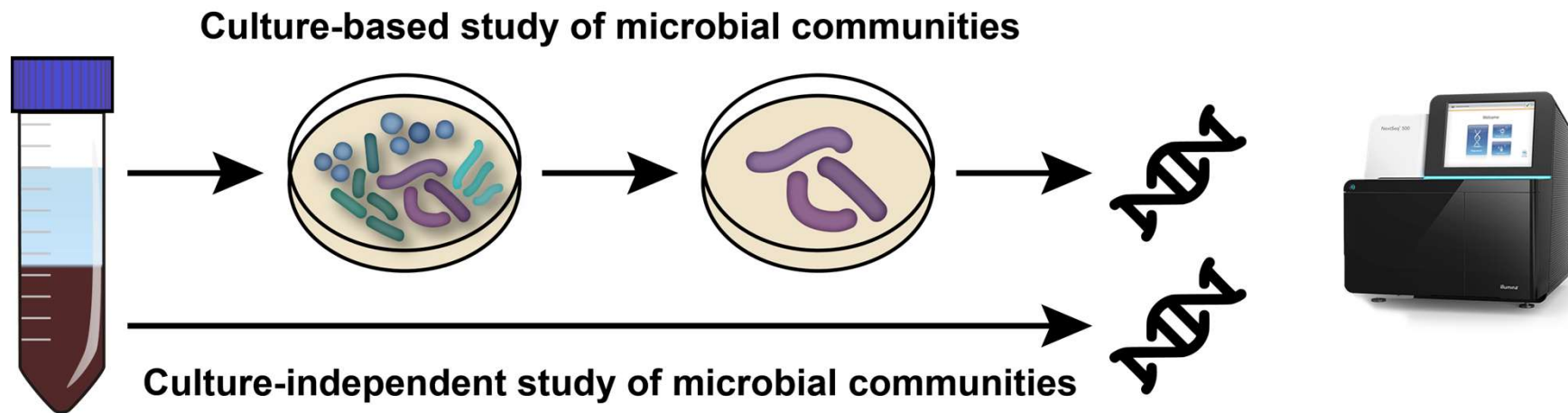


[alessio\\_mila](https://twitter.com/alessio_mila)

# Aims of this lecture

- Introduction of the **taxonomic profiling problem**
- **Conceptual overview** over approaches for taxonomic profiling
- **Sources of bias** in taxonomic profiling
- Introduction of the **mOTU profiling tool**
- **Comparative evaluation** of existing tools on community benchmarks
- How to account for **unknown species**, i.e. ones for which an isolate genome sequence is not available

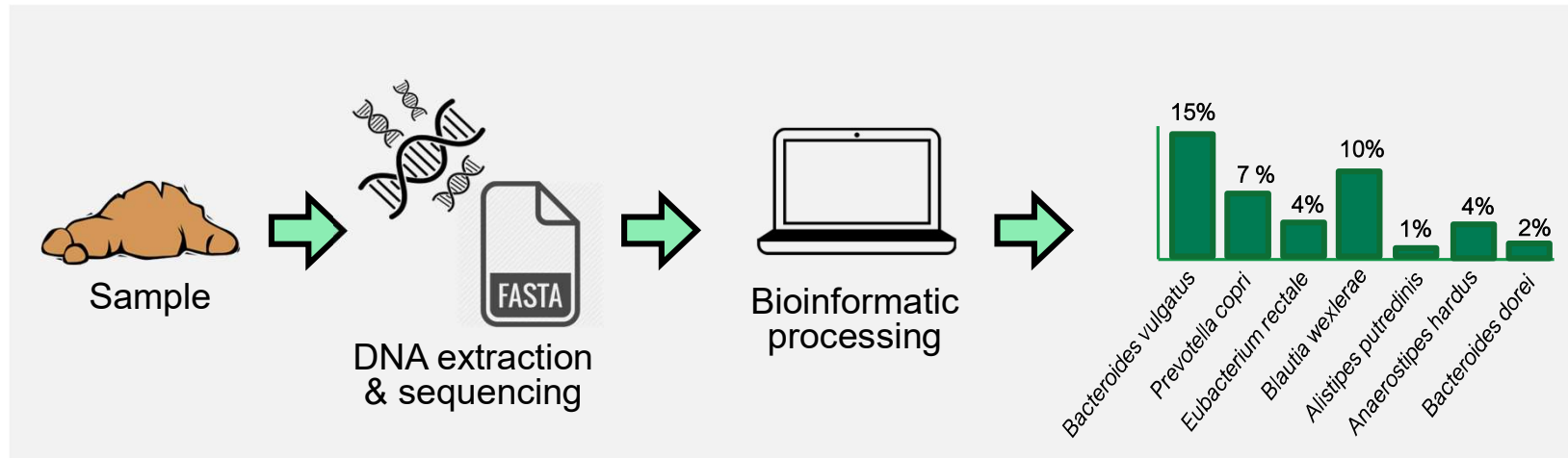
# Culture-independent sequencing of microbial communities



- Who's there? ➤ Taxonomic profiling
- Cave: DNA extraction and bioinformatics analysis are not standardized (technical biases)
- Key limitation: technology provides only snapshots, spatial information lost, causalities unclear

[Costea et al., *Nat. Biotechnol.*, 2017]

# Taxonomic profiling – what is it?



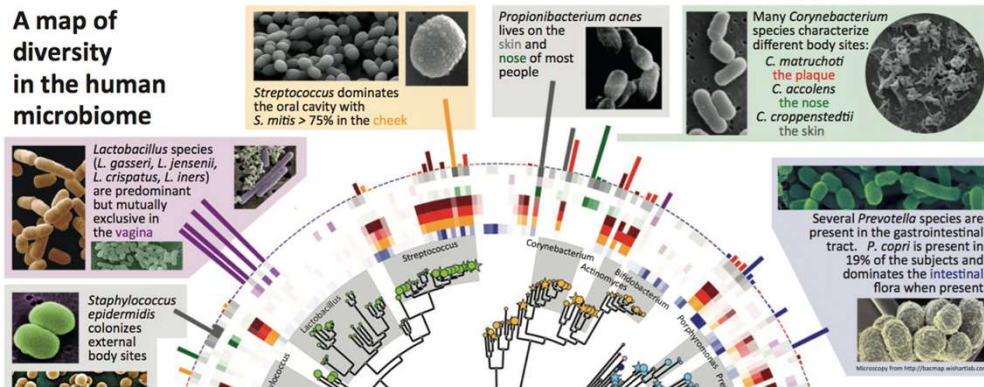
## Taxonomic Profiling:

Estimate relative cell counts in a microbiome sample from metagenomic sequencing

# Taxonomic profiling – why it is important?

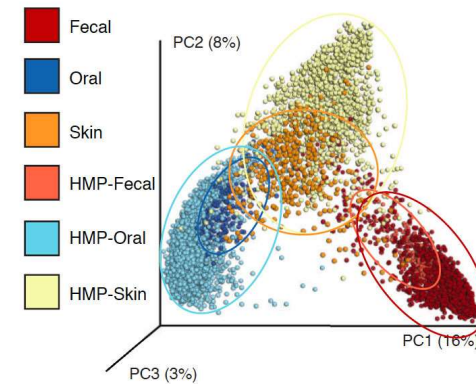
Taxonomic analysis is fundamental to the analysis of microbial communities

## Describing the microbial community under study



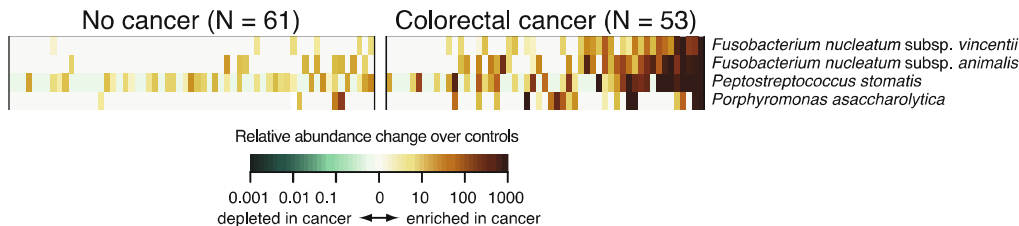
[Morgan et al., Trends in Genetics, 2013]

## Comparing different microbial communities



[McDonald et al., mSystems, 2018]

## Correlating environm. or host features to microbes



[Zeller et al., MSB, 2014]

## Comparing findings to literature

*Fusobacterium nucleatum* Contributes to the Carcinogenesis of Colorectal Cancer by Inducing Inflammation and Suppressing Host Immunity

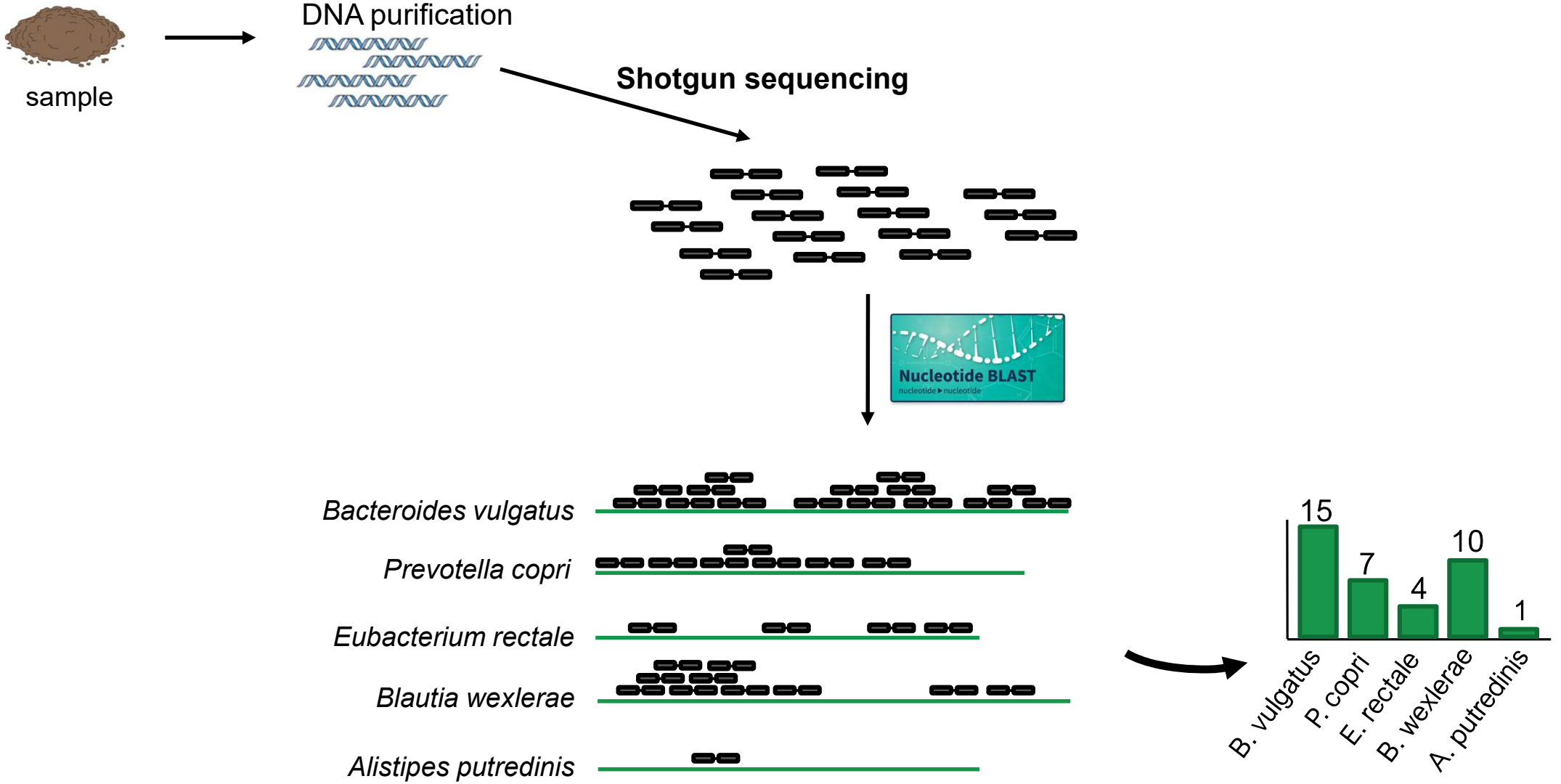
**RESEARCH**  
**CANCER**  
**Analysis of *Fusobacterium* persistence and antibiotic response in colorectal cancer**

Susan Brilman,<sup>1,2</sup> Chandira S. Pradamalla,<sup>1,2</sup> Ewa Sidnicka,<sup>1</sup> Thomas E. Clancy,<sup>4</sup> Xiaomeng Zhang,<sup>1,2</sup> Hana Cai,<sup>1,2</sup> Deena Neuberger,<sup>1</sup> Katherine Hwang,<sup>1</sup> Estelma Guerevara,<sup>1</sup> Timothy Nelson,<sup>1</sup> Otari Chipshachi,<sup>1</sup> Timothy Hagan,<sup>1</sup> Mark Walker,<sup>1</sup> Aruna Ramachandran,<sup>1</sup> Ingolia Doolado,<sup>1,2</sup> Garza Sierra,<sup>1</sup> Nurik Mink,<sup>1</sup> Stefania Landolfi,<sup>1</sup> Santiago Ramirez,<sup>1</sup> Celia,<sup>1</sup> Roberta Fasani,<sup>1</sup> Andrew J. Aguirre,<sup>1,2,3</sup> Kimmie Ng,<sup>1</sup> Elena Ezer,<sup>1</sup> Shuji Ogino,<sup>1,2</sup> Josep Taberner,<sup>1</sup> Charles S. Fuchs,<sup>1</sup> William C. Hahn,<sup>1,2,3</sup> Paulo Neufuss,<sup>1</sup> Matthew Meyerson<sup>1,2,3</sup>

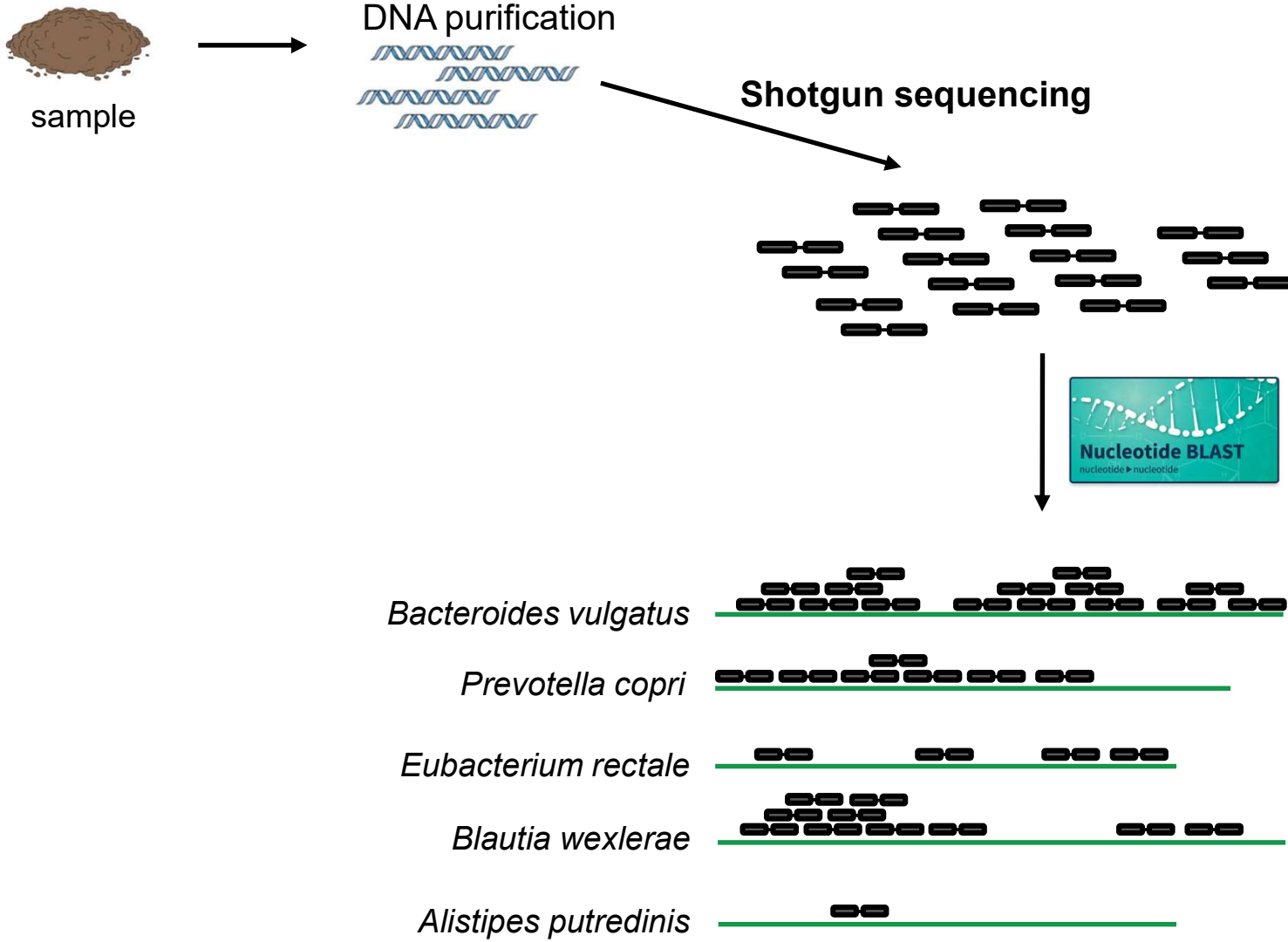
***Fusobacterium nucleatum* Promotes Colorectal Carcinogenesis by Modulating E-Cadherin/ $\beta$ -Catenin Signaling via its FadA Adhesin**

Mara Roxana Rubinsteyn,<sup>1,2</sup> Xiaowei Wang,<sup>1,2</sup> Wendy Lu,<sup>1,2</sup> Yujun Hao,<sup>1,2</sup> Guifang Cai,<sup>1</sup> and Yiping W. Han<sup>1,2,4\*</sup>  
<sup>1</sup>Department of Periodontics

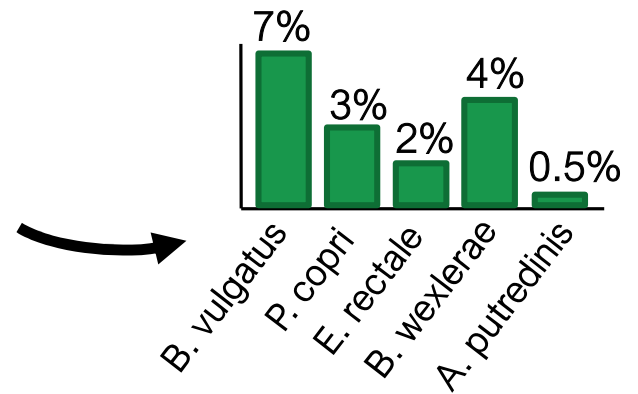
# Taxonomic profiling – how it is done?



# Taxonomic profiling – how it is done?

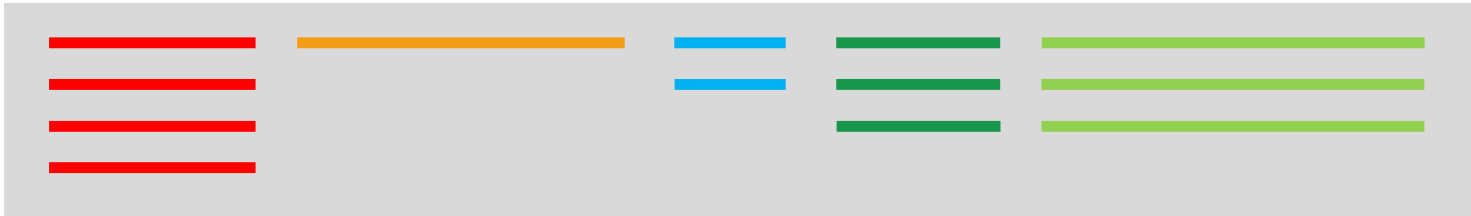


- Data is compositional

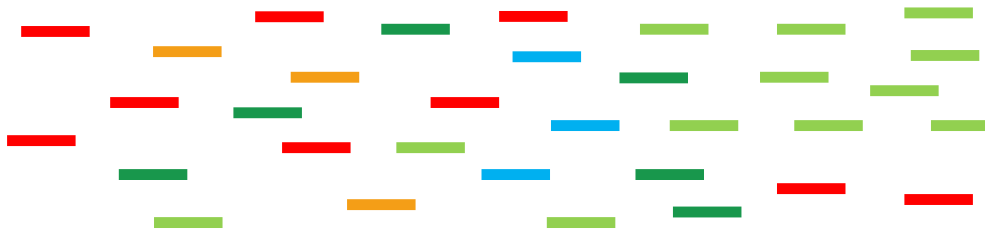


# Taxonomic profiling approaches – whole-genome mapping

Environmental sample



Shotgun sequencing

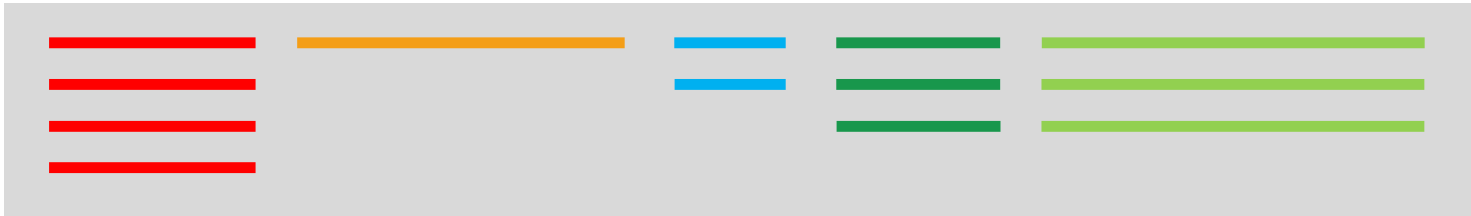


- DNA extraction bias
- sequencing biases
- sampling noise



# Taxonomic profiling approaches – whole-genome mapping

Environmental sample



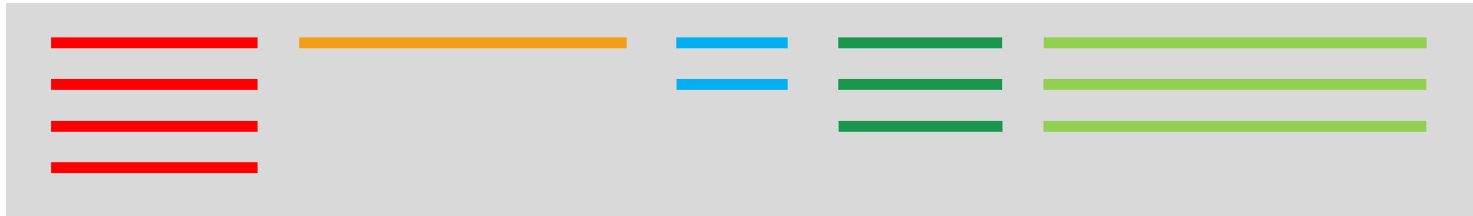
Shotgun sequencing



- DNA extraction bias
- sequencing biases
- sampling noise

# Taxonomic profiling approaches – whole-genome mapping

Environmental sample



Shotgun sequencing



- DNA extraction bias
- sequencing biases
- sampling noise

True taxonomic annotation



Estimated by whole-genome mapping



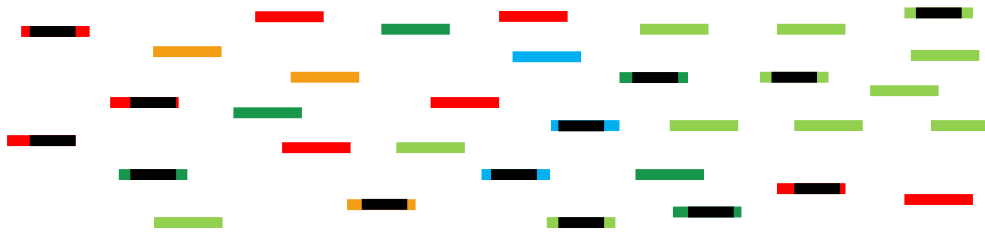
- genome size issue

# Taxonomic profiling approaches – marker gene mapping

Environmental sample



Shotgun sequencing



- DNA extraction bias
- sequencing biases
- sampling noise

# Taxonomic profiling approaches – marker gene mapping

Environmental sample



Shotgun sequencing



- DNA extraction bias
- sequencing biases
- sampling noise

# Taxonomic profiling approaches – marker gene mapping

Environmental sample



Shotgun sequencing



- DNA extraction bias
- sequencing biases
- sampling noise

# Taxonomic profiling approaches – marker gene mapping

Environmental sample



Shotgun sequencing



- DNA extraction bias
- sequencing biases
- sampling noise

True taxonomic annotation



- genome size issue

# Taxonomic profiling – mapping reads to genomes

Environmental sample



Shotgun sequencing



# Taxonomic profiling – incomplete reference databases

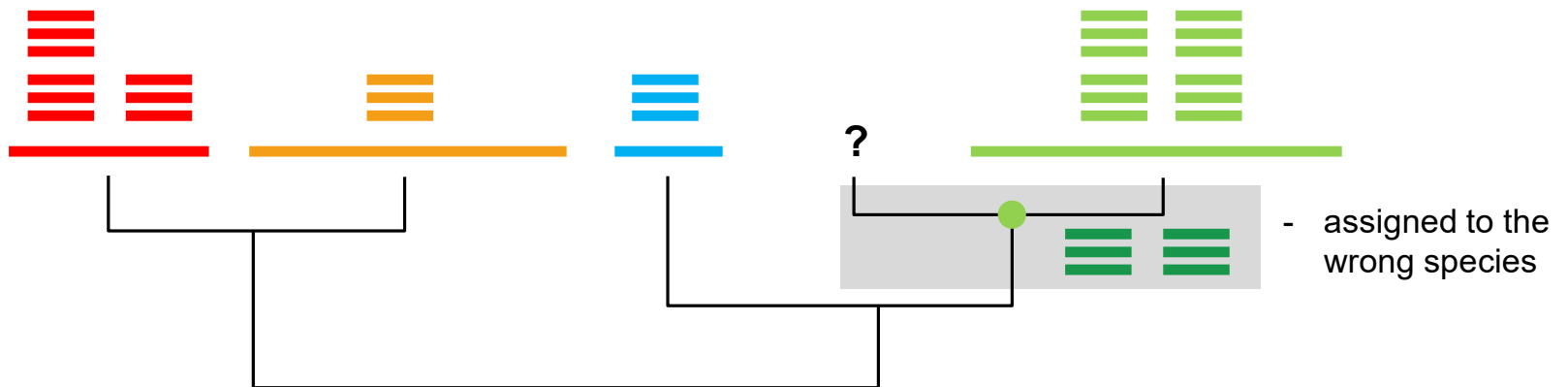
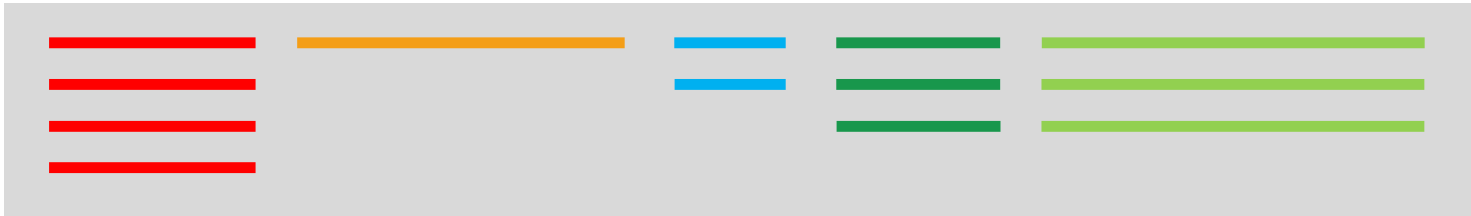
Environmental sample





# Taxonomic profiling – incomplete reference databases

Environmental sample



# Taxonomic profiling – incomplete reference databases

Environmental sample



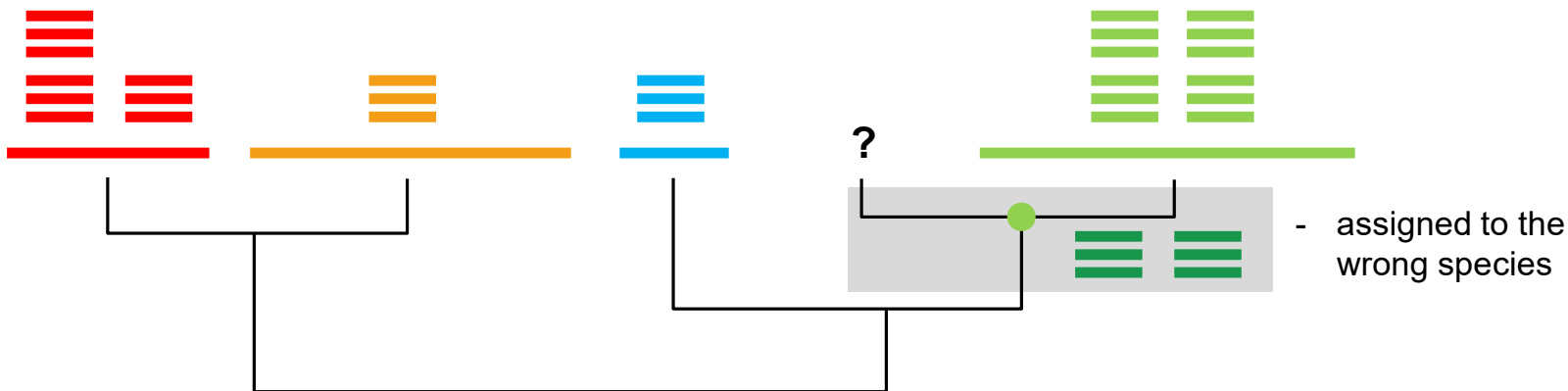
True taxonomic annotation



Estimated when dark green is missing



(ignore genome size issue)



# Taxonomic profiling – incomplete reference databases

Environmental sample



- ignore the reads



# Taxonomic profiling – incomplete reference databases

Environmental sample



True taxonomic annotation



Estimated when dark green is missing

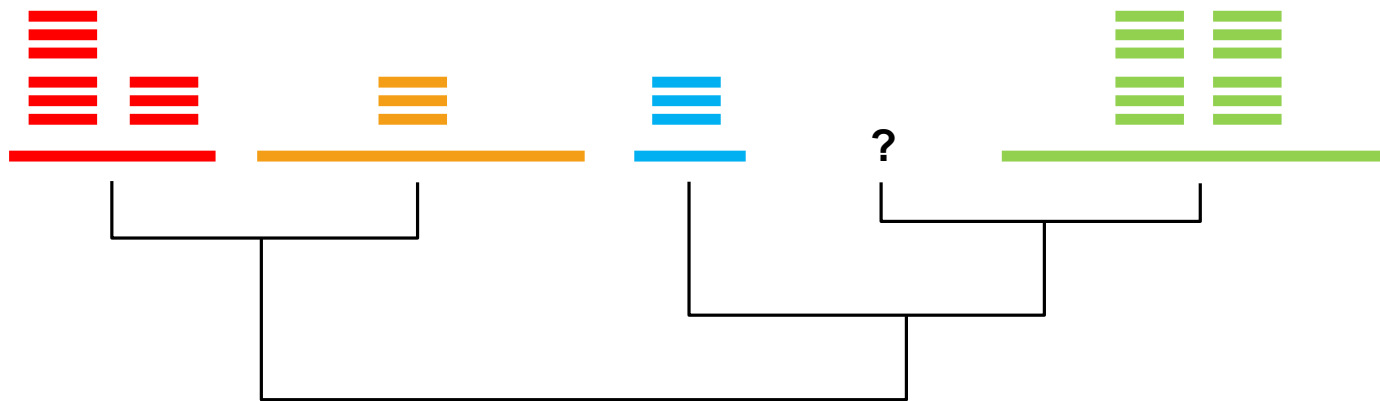


- ignore the reads

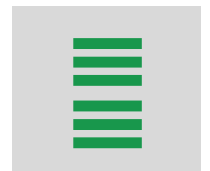


# Taxonomic profiling – incomplete reference databases

Environmental sample



Globally unassigned



# Taxonomic profiling – incomplete reference databases

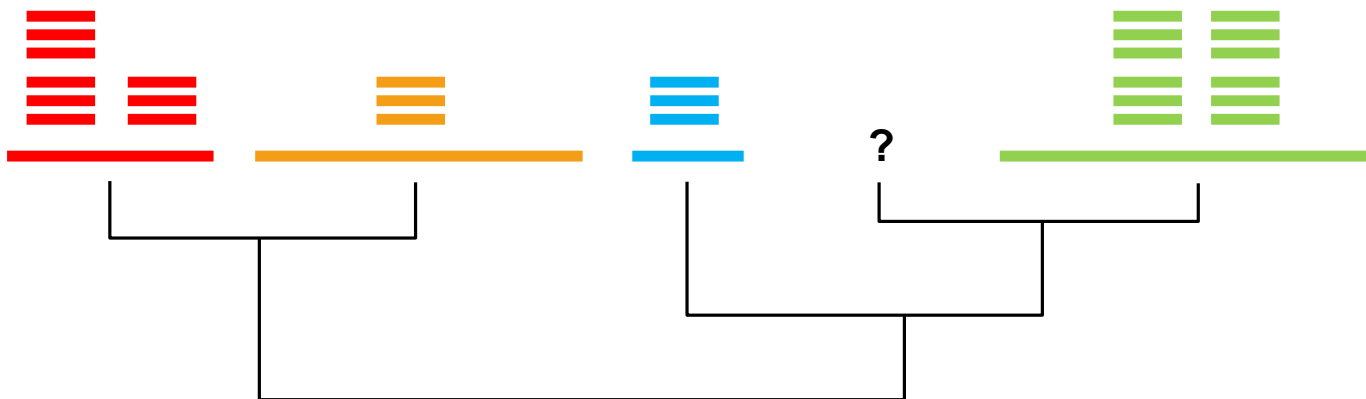
Environmental sample



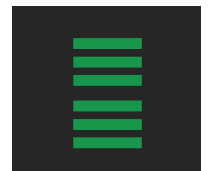
True taxonomic annotation



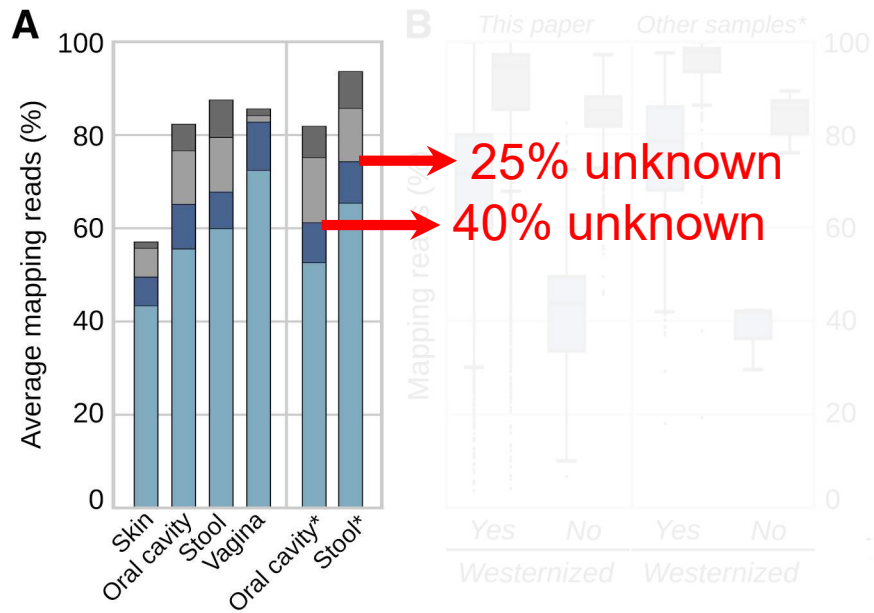
Estimated when dark green is missing



Globally unassigned



# How much is unknown?



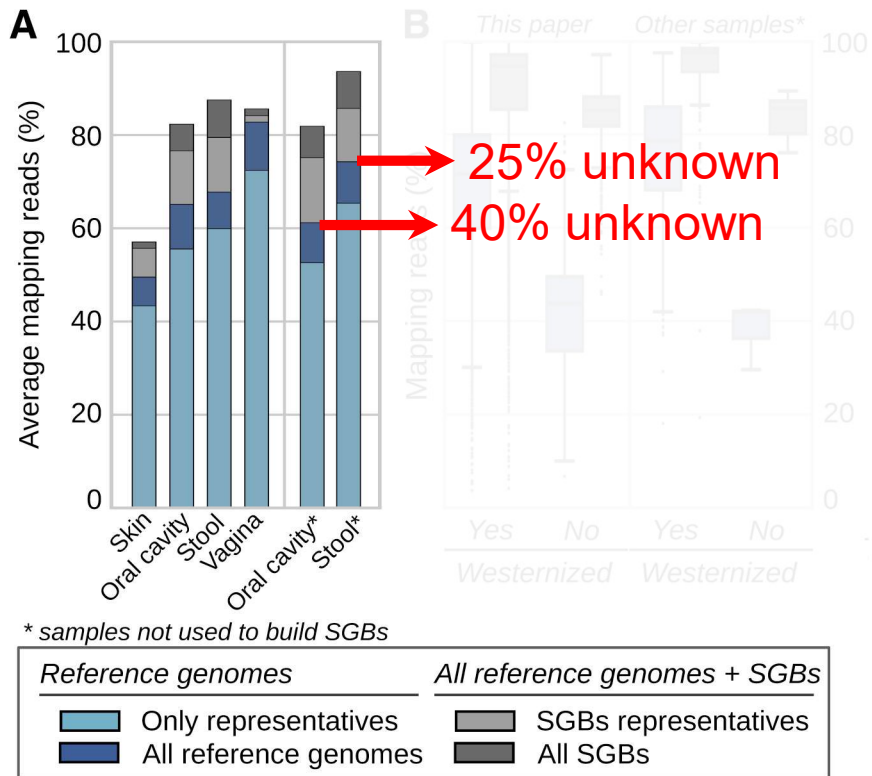
25% unknown  
40% unknown

\* samples not used to build SGBs

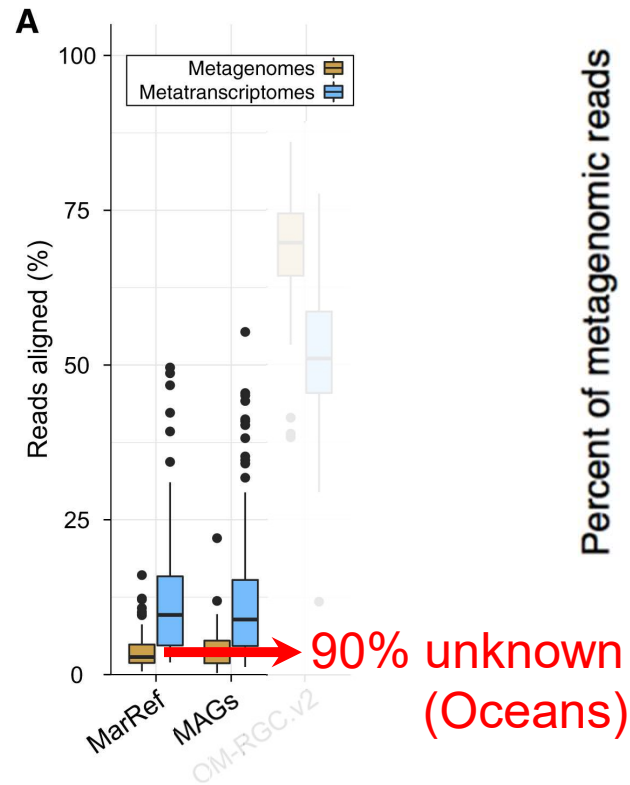
Reference genomes	All reference genomes + SGBs
Only representatives	SGBs representatives
All reference genomes	All SGBs

[Pasolli et al., Cell, 2019]

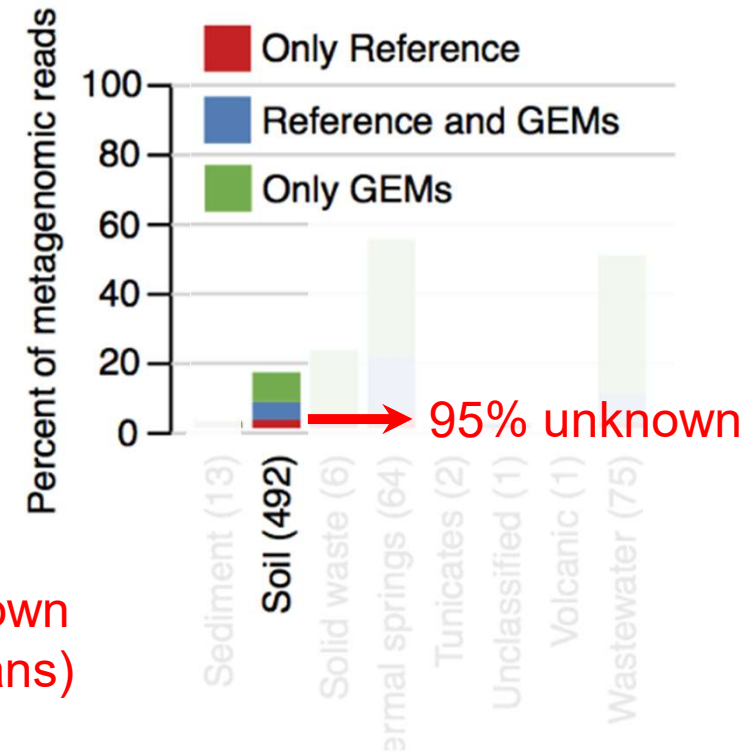
# How much is unknown?



[Pasolli et al., *Cell*, 2019]



[Salazar et al., *Cell*, 2019]



[Nayfach et al., *Nat. Biotechnol.*, 2021]



# Taxonomic profiling approaches – marker gene mapping

Environmental sample



Shotgun sequencing



- DNA extraction bias
- sequencing biases
- sampling noise

True taxonomic annotation



- genome size issue

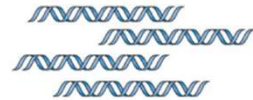
# Taxonomic profiling approaches – 16S rRNA amplicon



sample



DNA purification



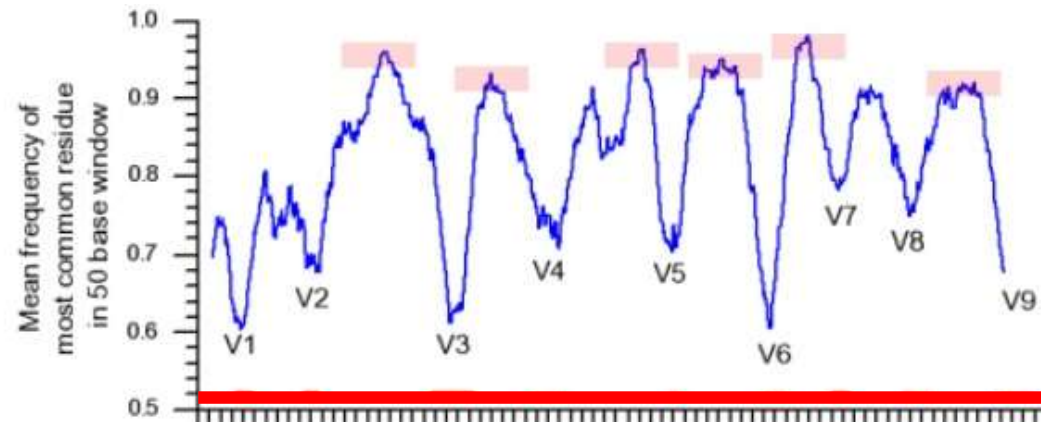
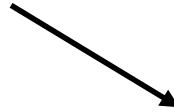
# Taxonomic profiling approaches – 16S rRNA amplicon



sample



DNA purification



potential PCR primer sites

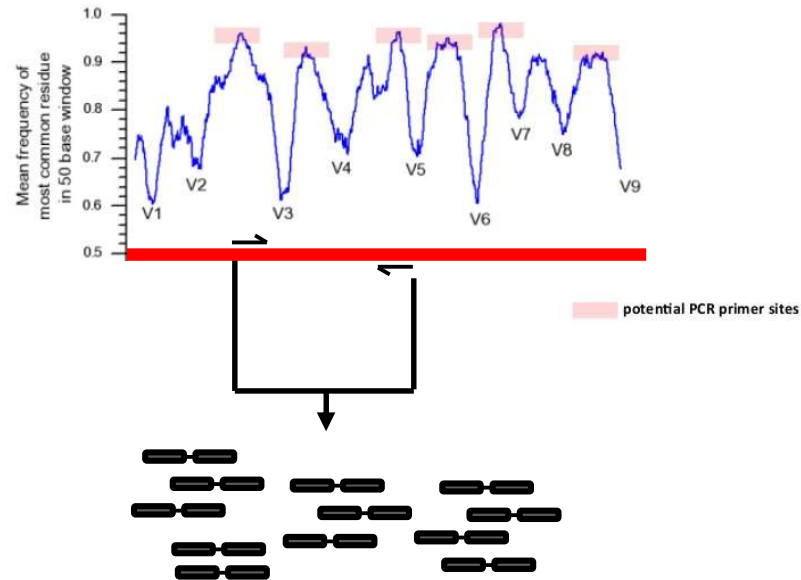
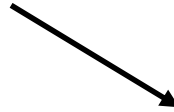
# Taxonomic profiling approaches – 16S rRNA amplicon



sample



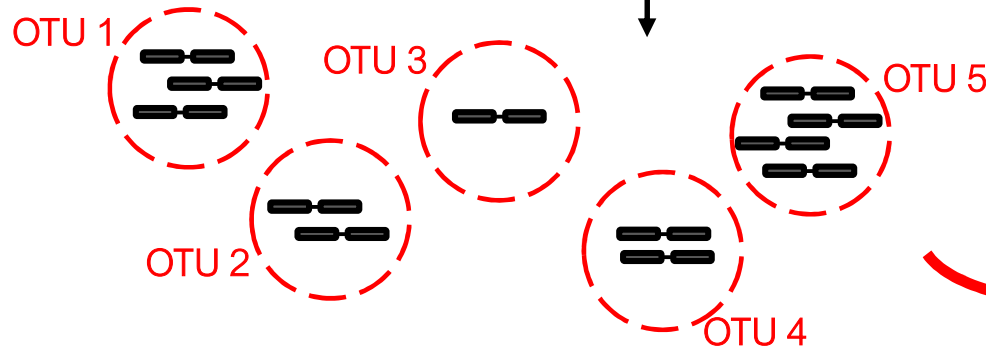
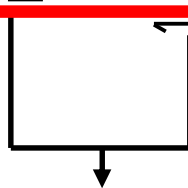
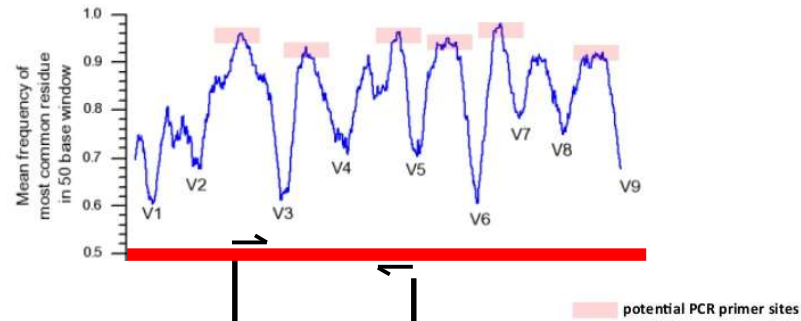
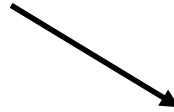
DNA purification



# Taxonomic profiling approaches – 16S rRNA amplicon

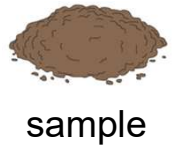


DNA purification

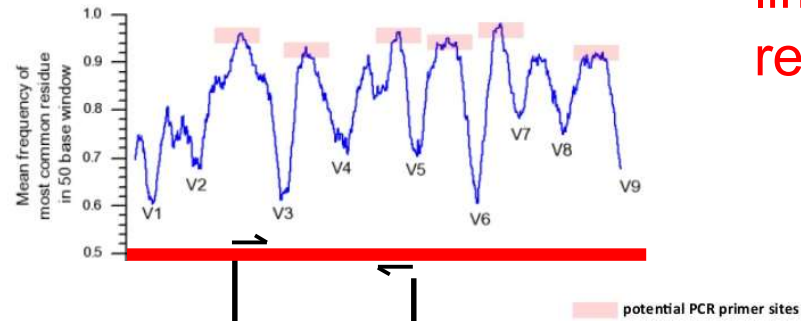
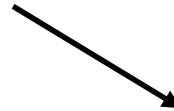
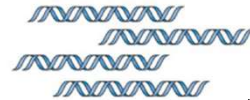


OTU = „operational taxonomic unit“

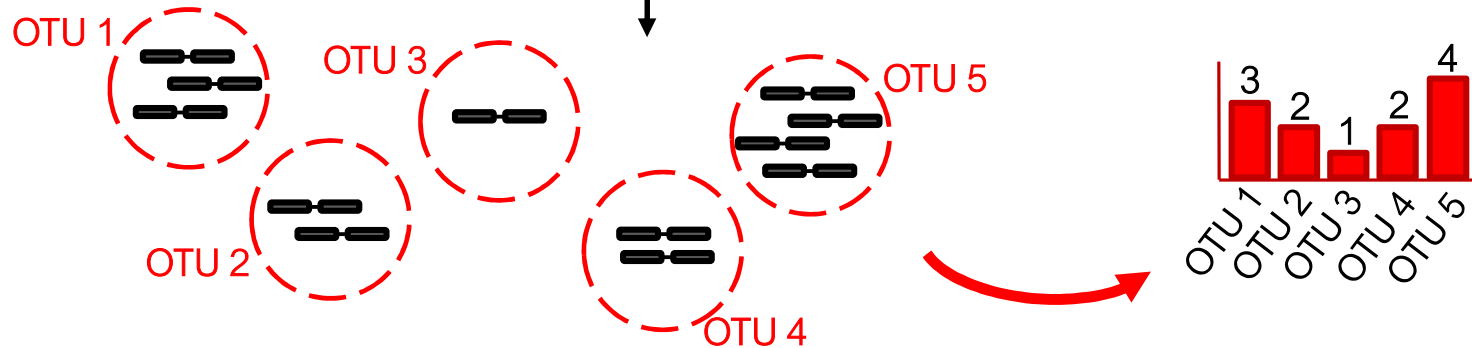
# Taxonomic profiling approaches – 16S rRNA amplicon



DNA purification



**! WARNING:**  
The OTUs have a length of 250/400 which results in limited taxonomic resolution



OTU = „operational taxonomic unit“



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Published: 20 October 2013

### Metagenomic species profiling using universal phylogenetic marker genes

[Shinichi Sunagawa](#), [Daniel R Mende](#), [Georg Zeller](#), [Fernando Izquierdo-Carrasco](#), [Simon A Berger](#), [Jens Roat Kultima](#), [Luis Pedro Coelho](#), [Manimozhayan Arumugam](#), [Julien Tap](#), [Henrik Bjørn Nielsen](#), [Simon Rasmussen](#), [Søren Brunak](#), [Oluf Pedersen](#), [Francisco Guarner](#), [Willem M de Vos](#), [Jun Wang](#), [Junhua Li](#), [Joël Doré](#), [S Dusko Ehrlich](#), [Alexandros Stamatakis](#) & [Peer Bork](#) [✉](#)

*Nature Methods* **10**, 1196–1199 (2013) | [Cite this article](#)

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Article | [Open Access](#) | Published: 04 March 2019

### Microbial abundance, activity and population genomic profiling with mOTUs2

[Alessio Milanese](#), [Daniel R Mende](#), [Lucas Paoli](#), [Guillem Salazar](#), [Hans-Joachim Ruscheweyh](#), [Miquelangel Cuenca](#), [Pascal Hingamp](#), [Renato Alves](#), [Paul I Costea](#), [Luis Pedro Coelho](#), [Thomas S. B. Schmidt](#), [Alexandre Almeida](#), [Alex L Mitchell](#), [Robert D. Finn](#), [Jaime Huerta-Cepas](#), [Peer Bork](#), [Georg Zeller](#) [✉](#) & [Shinichi Sunagawa](#) [✉](#)

*Nature Communications* **10**, Article number: 1014 (2019) | [Cite this article](#)

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### mOTUs: Profiling Taxonomic Composition, Transcriptional Activity and Strain Populations of Microbial Communities

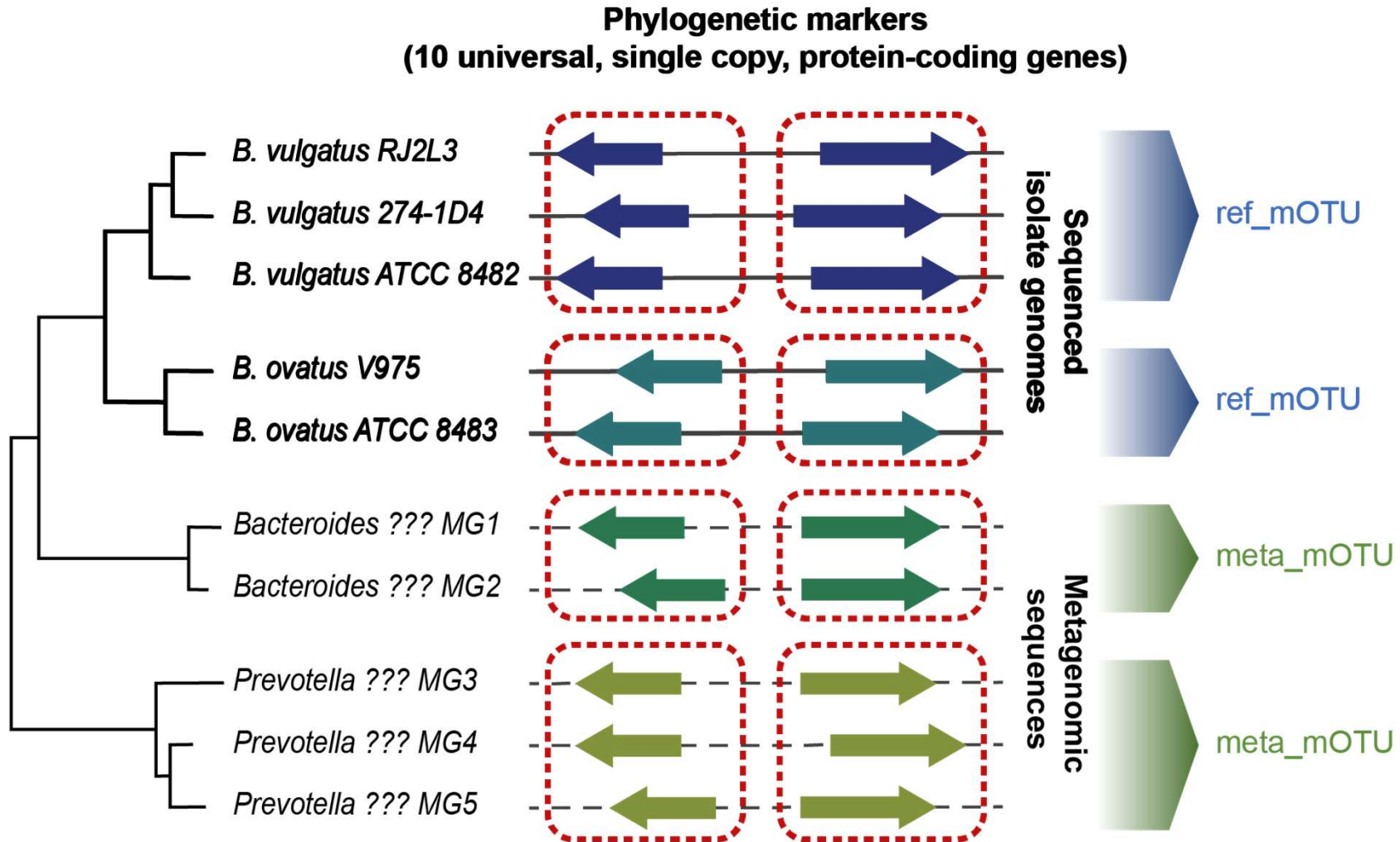
[Hans-Joachim Ruscheweyh](#), [Alessio Milanese](#), [Lucas Paoli](#), [Anna Sintsova](#), [Daniel R. Mende](#), [Georg Zeller](#) [✉](#), [Shinichi Sunagawa](#) [✉](#)

First published: 13 August 2021 | <https://doi.org/10.1002/cpz1.218>

[sfx.ethz.ch](#)

# The mOTUs framework – DB construction

[Ciccarelli et al. *Science* 2006]  
[Sunagawa et al. *Nat. Methods* 2013]  
[Milanese et al. *Nat. Commun.* 2019]

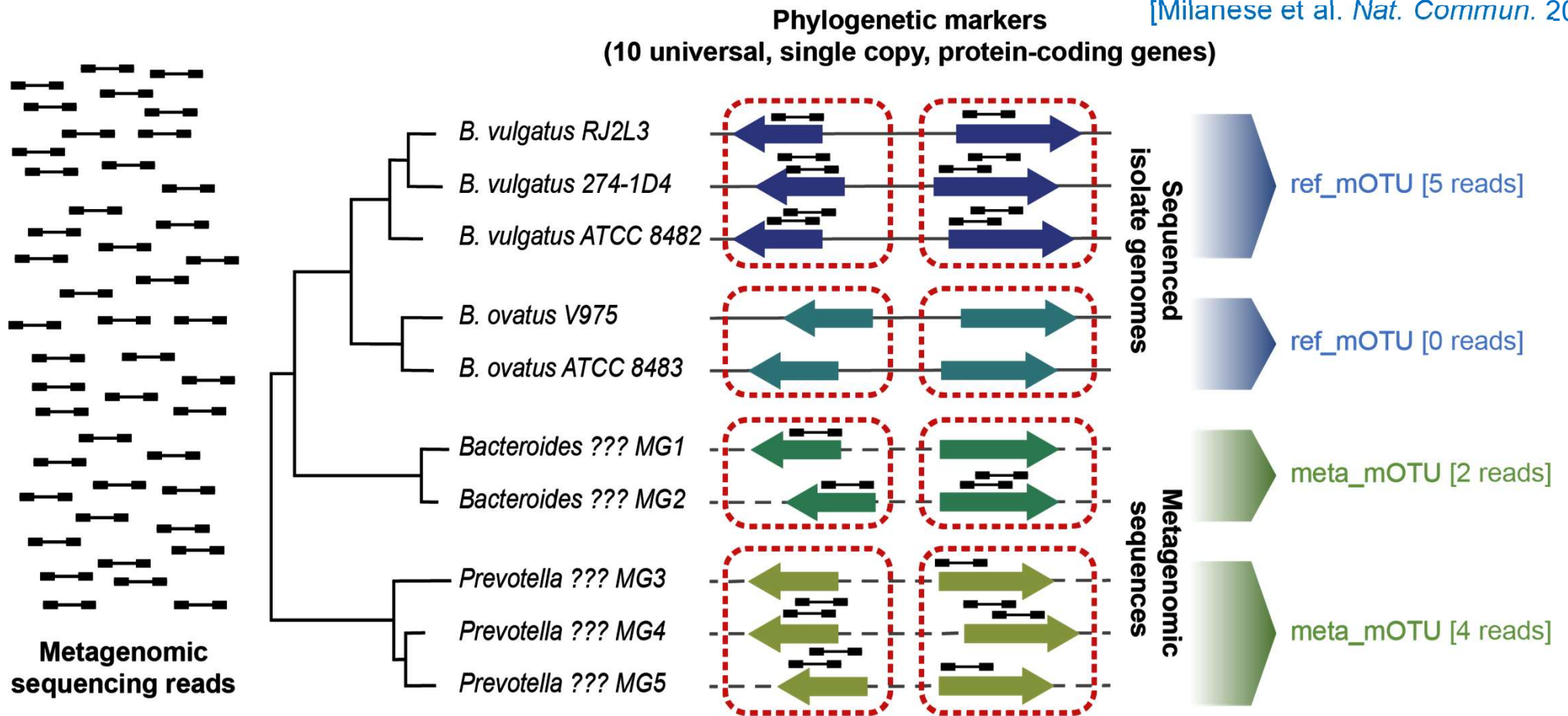




# The mOTUs framework – mapping / profiling

[Sunagawa et al. *Nat. Methods* 2013]

[Milanese et al. *Nat. Commun.* 2019]



Metagenomic  
sequencing reads

*B. vulgatus* RJ2L3  
*B. vulgatus* 274-1D4  
*B. vulgatus* ATCC 8482  
*B. ovatus* V975  
*B. ovatus* ATCC 8483  
*Bacteroides* ??? MG1  
*Bacteroides* ??? MG2  
*Prevotella* ??? MG3  
*Prevotella* ??? MG4  
*Prevotella* ??? MG5

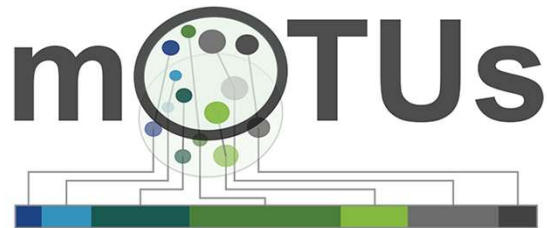
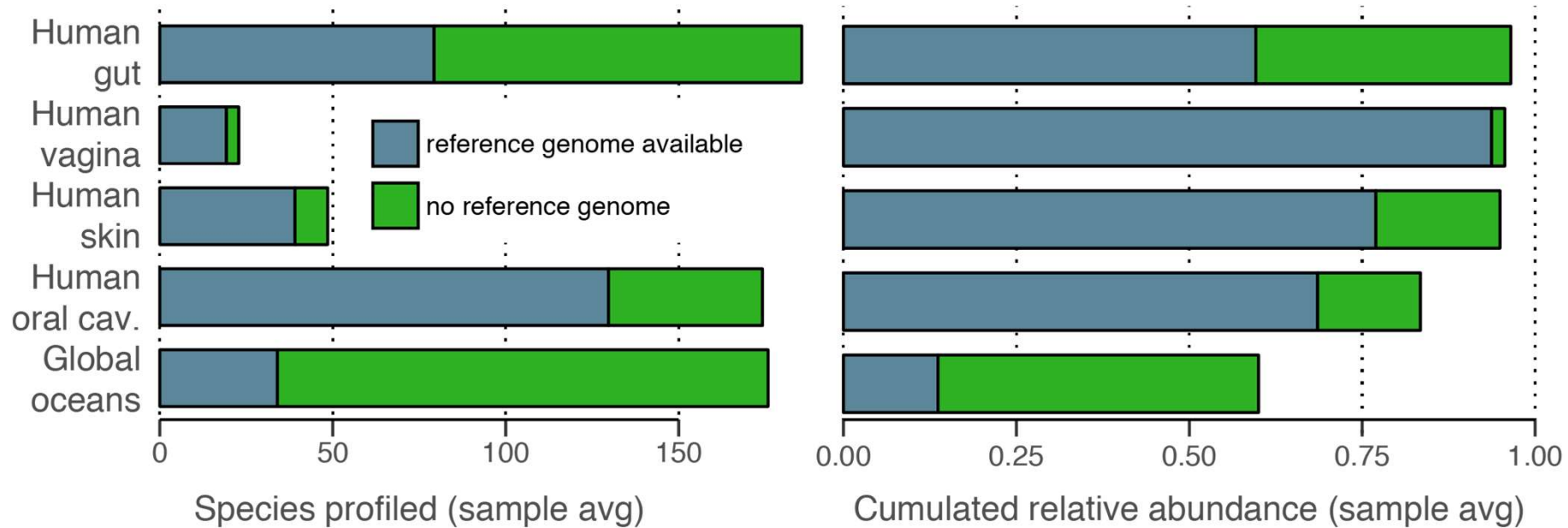
Sequenced  
isolate genomes  
Metagenomic  
sequences

ref\_mOTU [5 reads]  
ref\_mOTU [0 reads]  
meta\_mOTU [2 reads]  
meta\_mOTU [4 reads]

**Co-abundance binning**

**Break till 14:20**

# High-accuracy profiling of uncharacterized microbial species



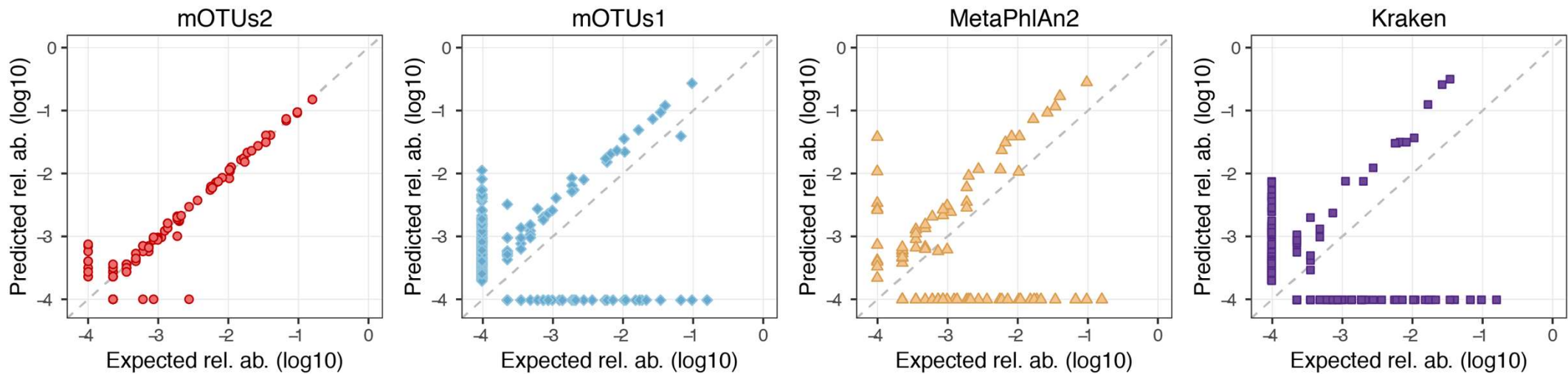
- High recall (non-reference species)
- High accuracy
- High resolution (up to subspecies)
- Also useful for meta-transcriptomics

[Milanese et al., *Nat. Commun.* 2019]

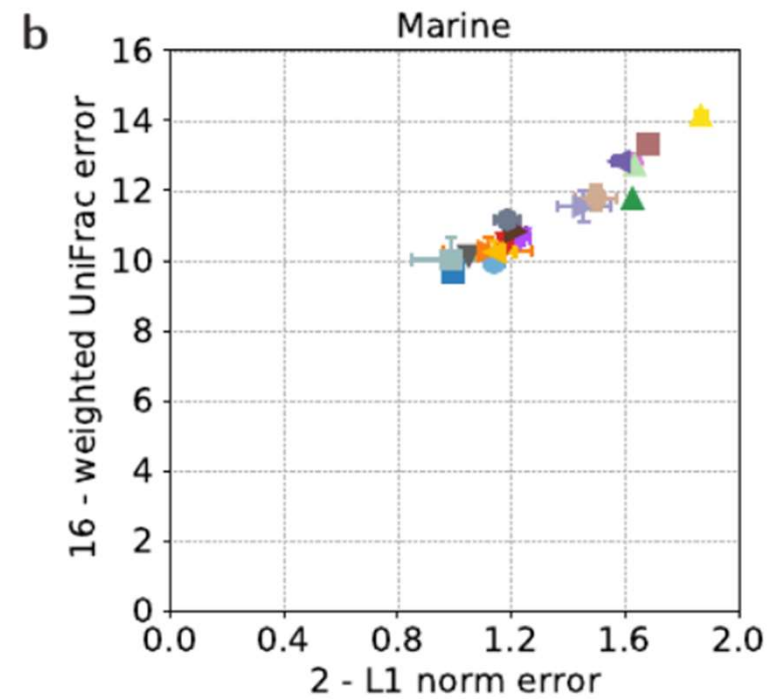
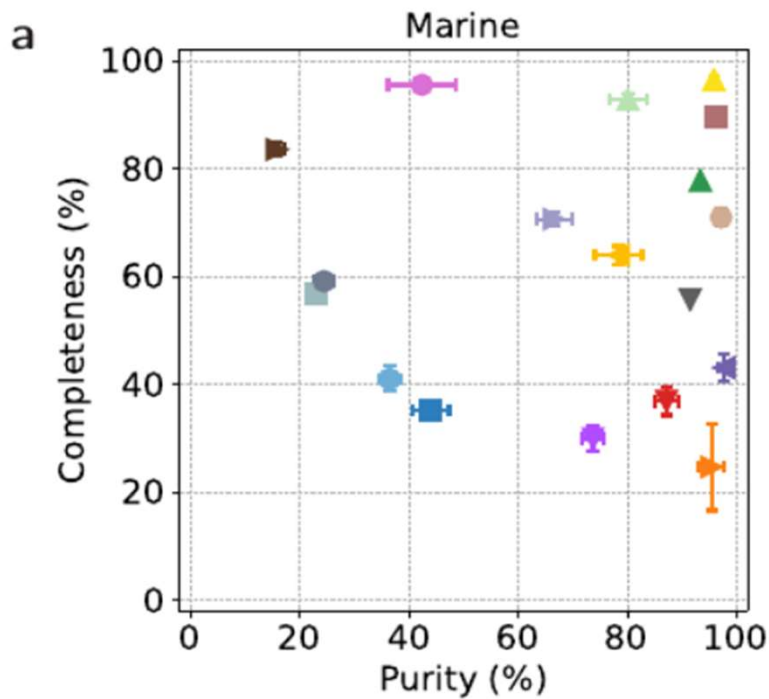
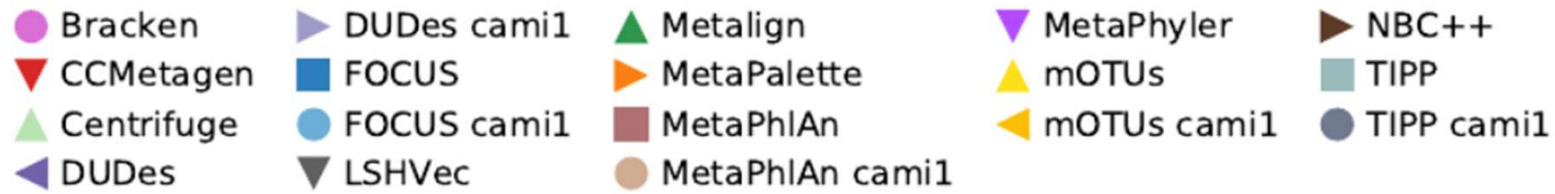
# Improved quantification owing to comprehensive DB

**Missing species** in the reference database lead to **overestimating the abundance of known taxa**, i.e. to biased quantification.

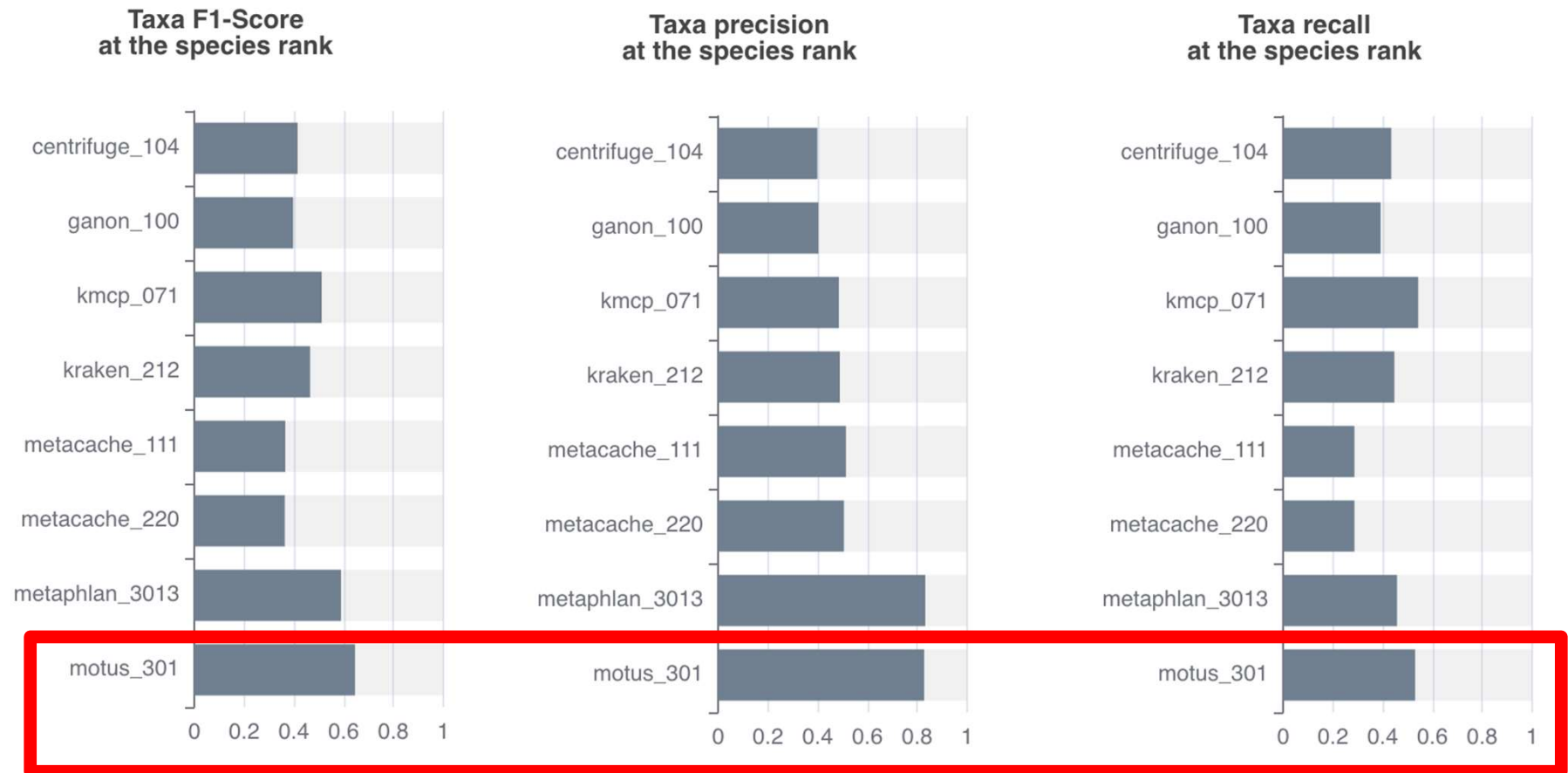
Simulations based on metagenome-assembled genomes



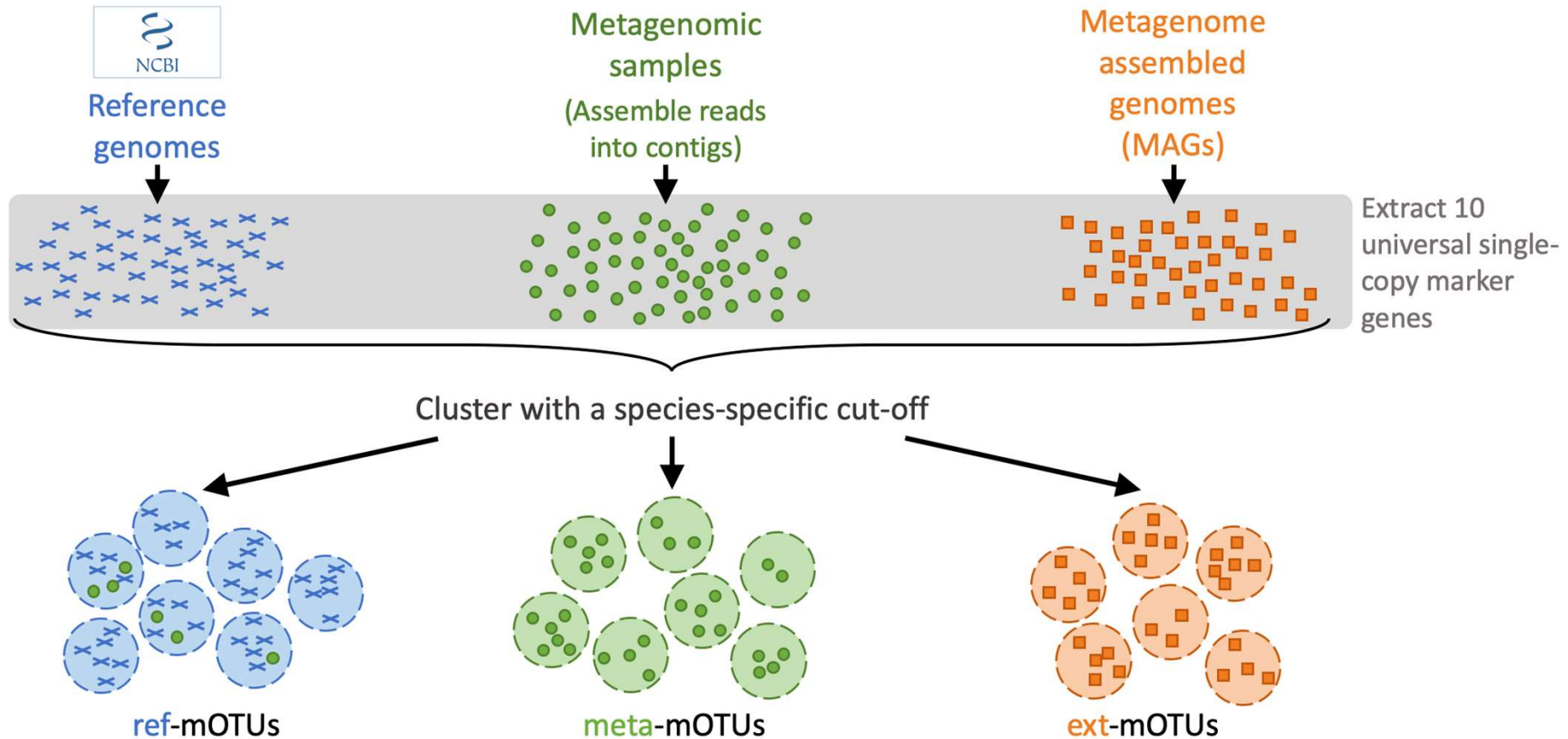
# High-accuracy profiling as evaluated by an independent benchmark - CAMI



# High-accuracy profiling as evaluated by an independent benchmark - LEMMI



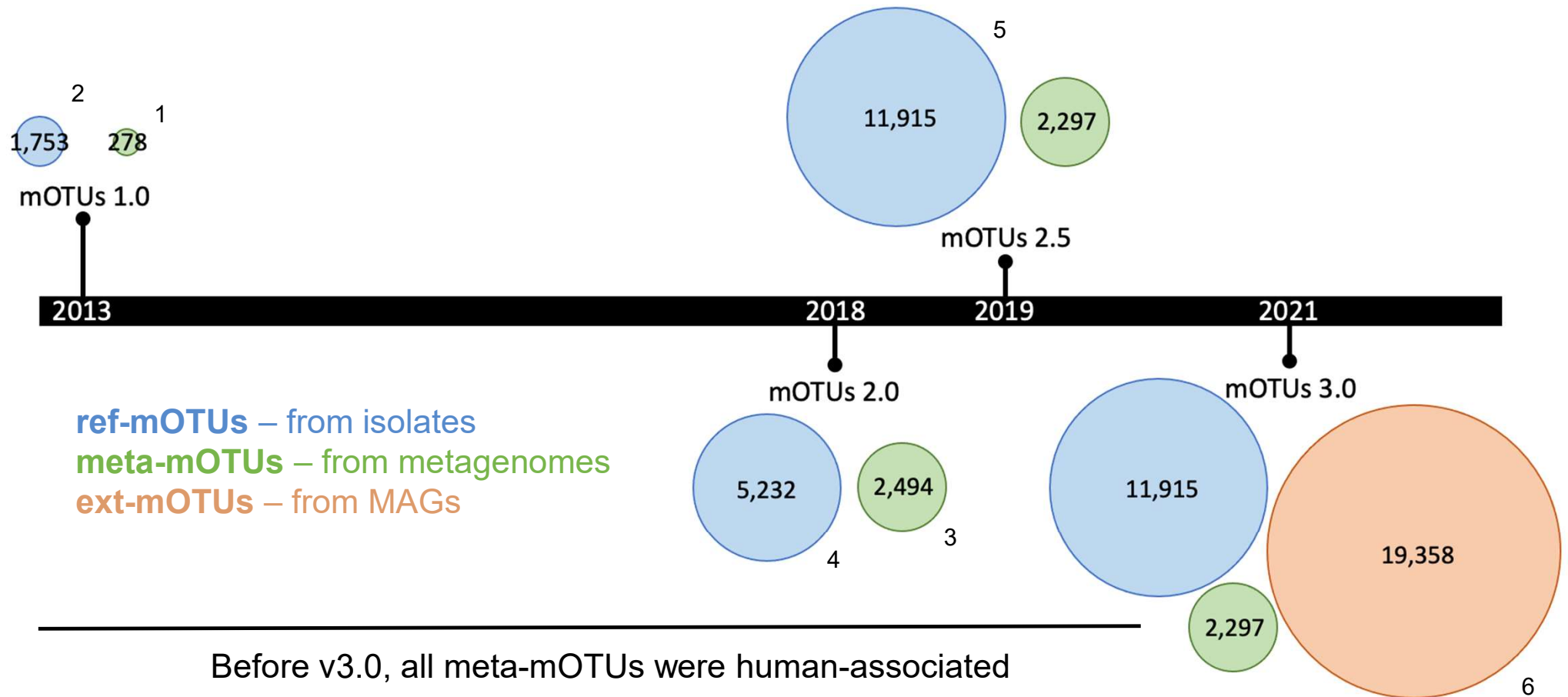
# Incorporation of MAGs into the mOTUs3 database



MAG-derived mOTUs are called ext\_mOTUs



# Improvement of scope in mOTUs since first version



1. [Sunagawa et al., *Nat. Methods* 2013]

2. [Mende et al. *Nat. Methods* 2013]

3. [Milanese et al., *Nat. Commun.* 2019]

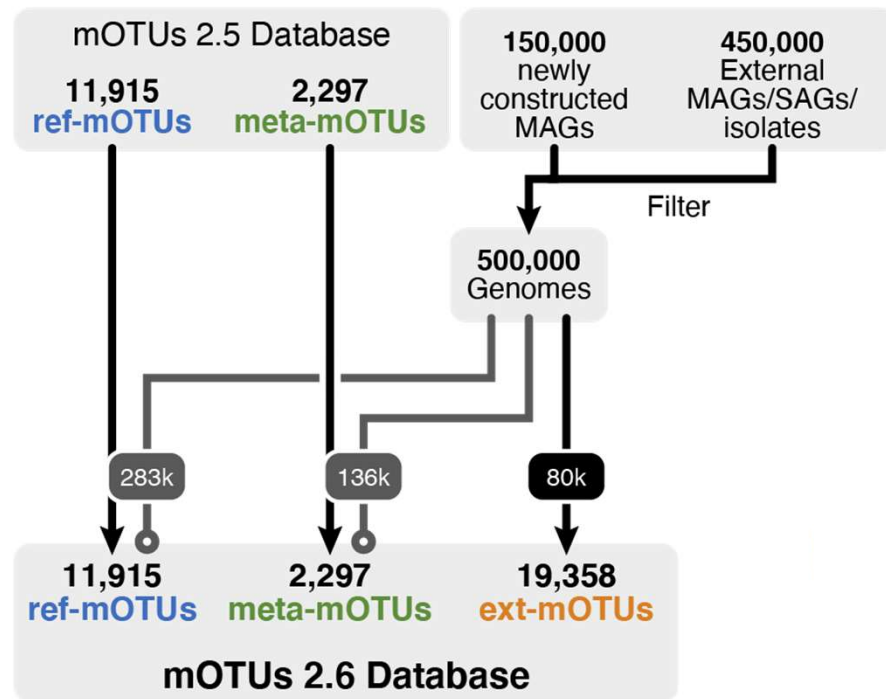
4. [Mende et al., *Nucleic Acids Res.* 2017]

5. [Mende et al., *Nucleic Acids Res.* 2020]

6. [Ruscheweyh, Milanese et al. *bioRxiv* 2021]

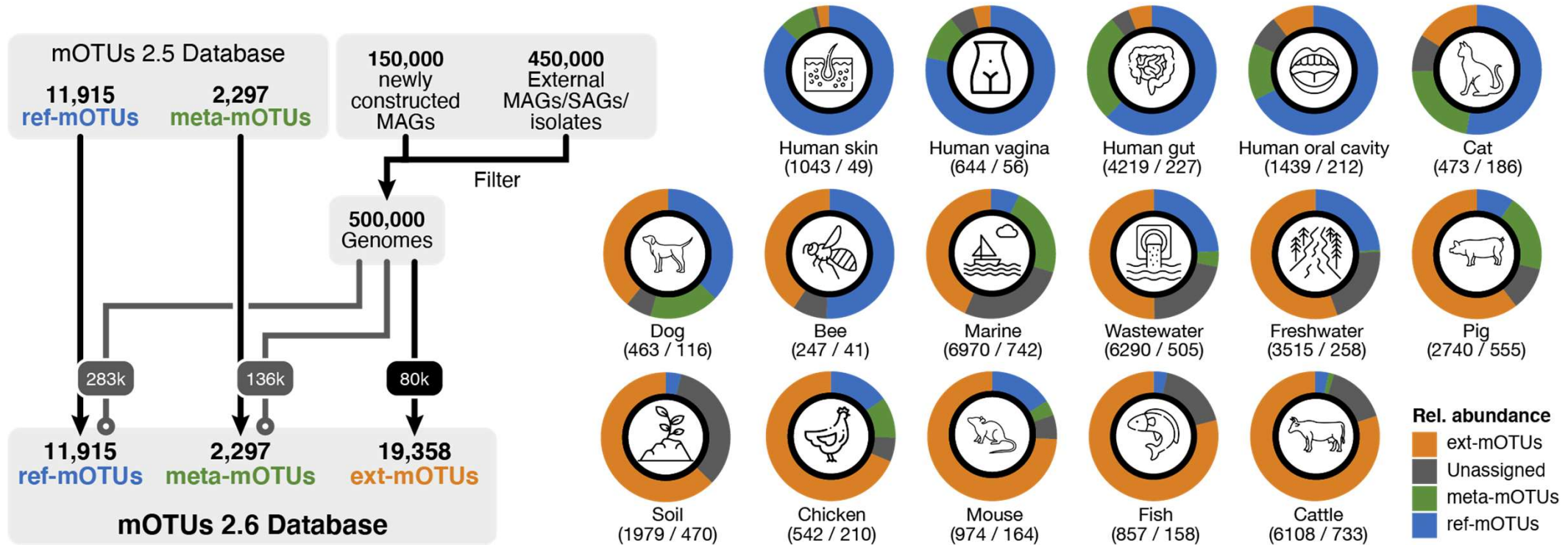


# mOTUs3 – database extension by marker genes from metagenome-assembled genomes (>500,000 MAGs)



**How do you map MAGs to mOTUs?**

# mOTUs3 – database extension by marker genes from metagenome-assembled genomes (>500,000 MAGs)



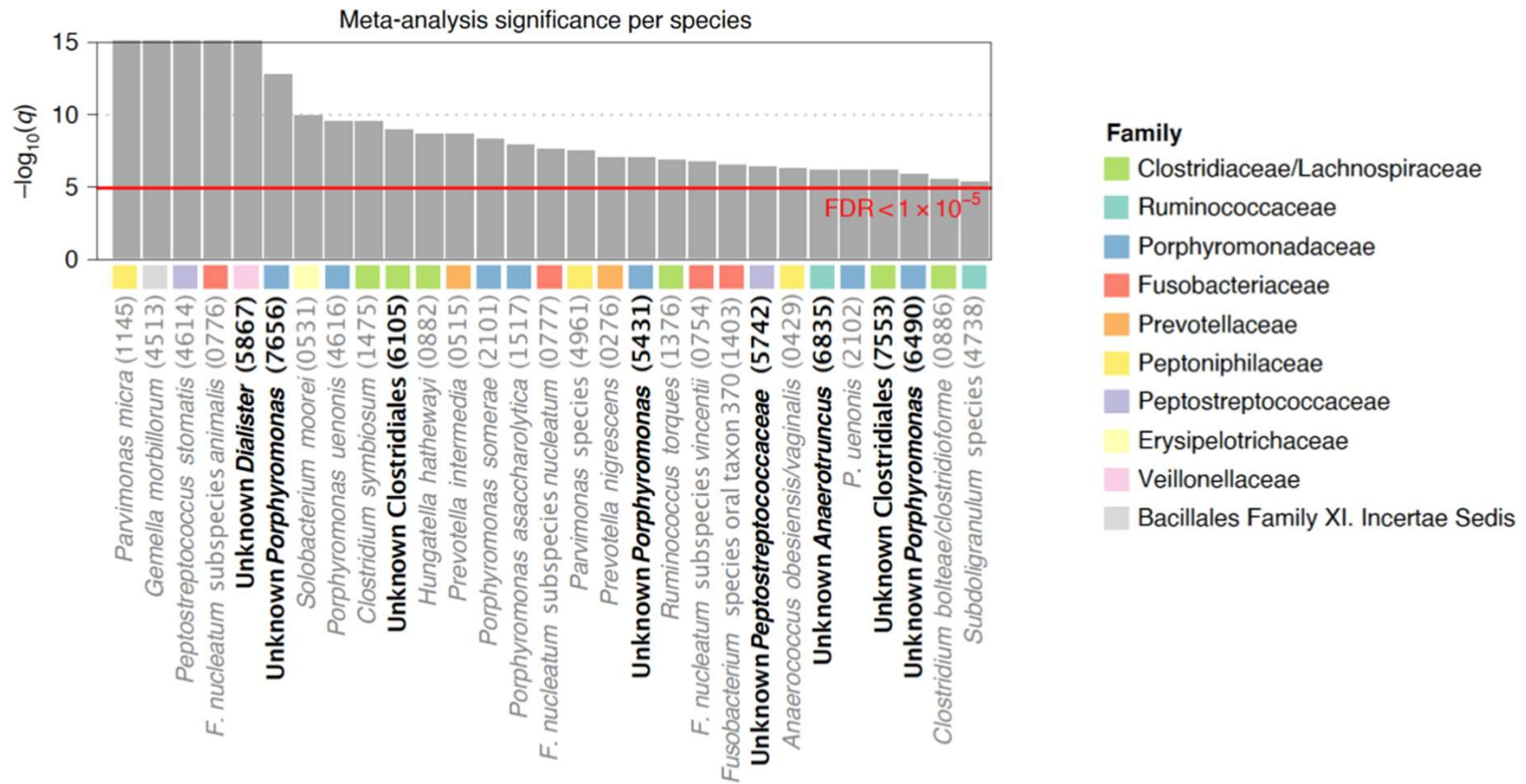
Enables profiling an unprecedented diversity of prokaryotes (33,570 species) across many environments.

[motu-tool.org](http://motu-tool.org)

[Ruschewey, Milanese et al., bioRxiv 2021]

# What is the added value of using mOTUs?

Since we are able to profile unknown species, we can study associations of unknown species to disease

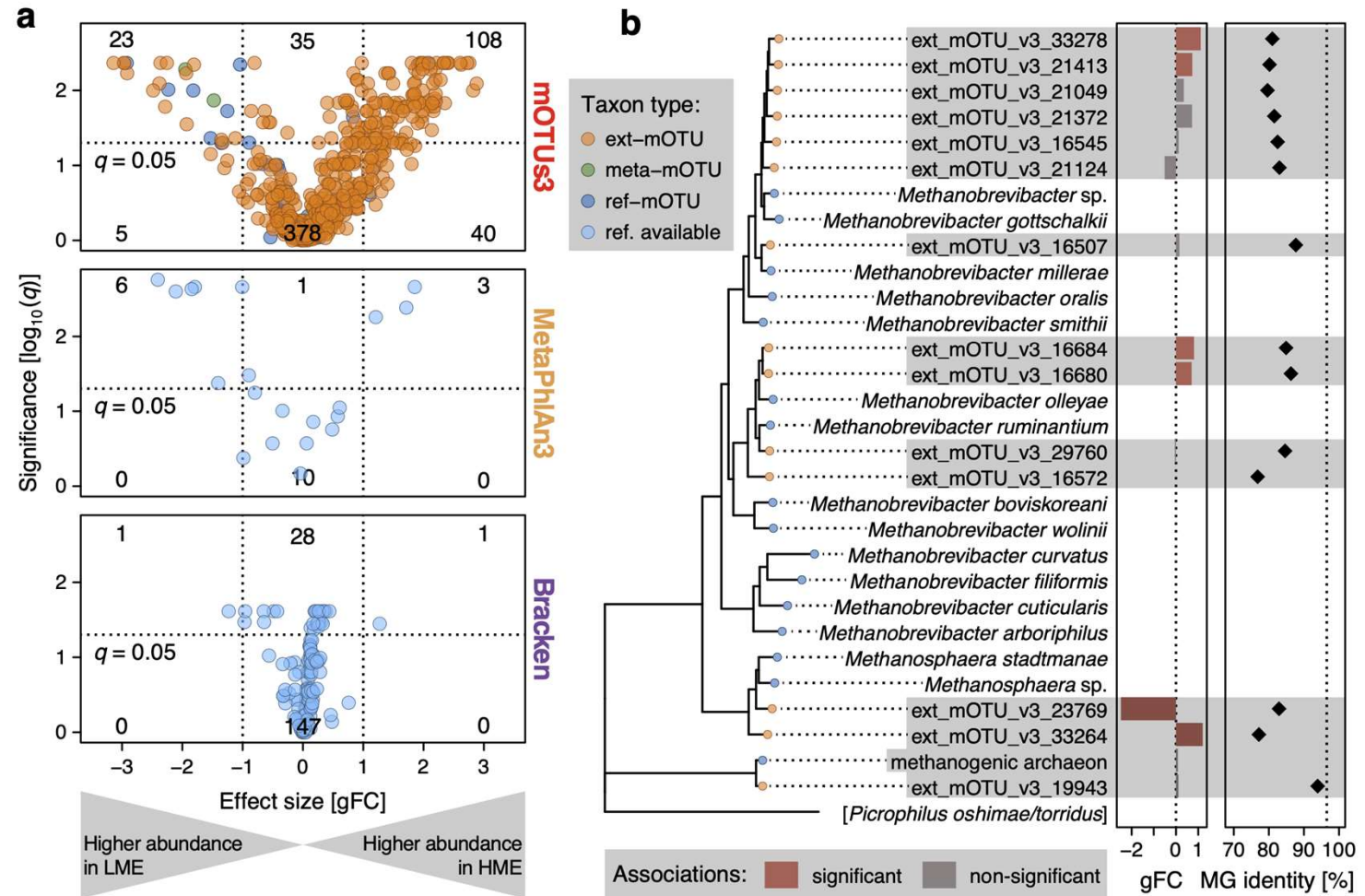


# What is the added value of using mOTUs?

- To demonstrate the utility of mOTUs3, we reanalysed rumen metagenomes from high- and low-methane emitting (HME and LME) sheep.

**LME** = low-methane emitting sheep

**HME** = high-methane emitting sheep

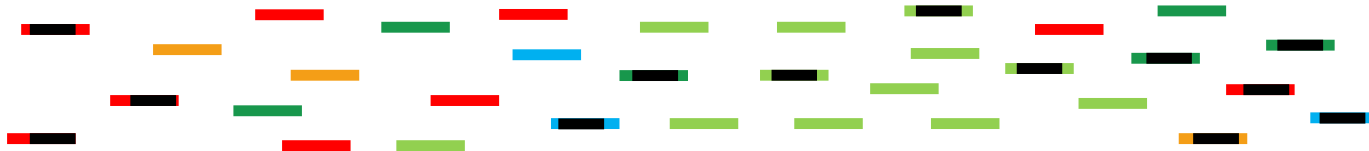


# Functional profiling

Environmental sample



Shotgun sequencing



Taxonomic profiling

- Universal marker gene reads can be used for taxonomic profiling

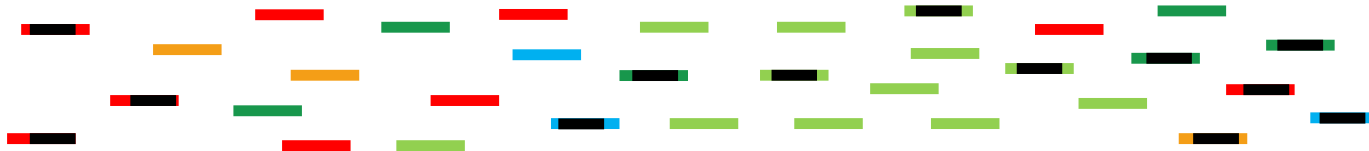


# Functional profiling

Environmental sample



Shotgun sequencing



Taxonomic profiling

- Universal marker gene reads can be used for taxonomic profiling

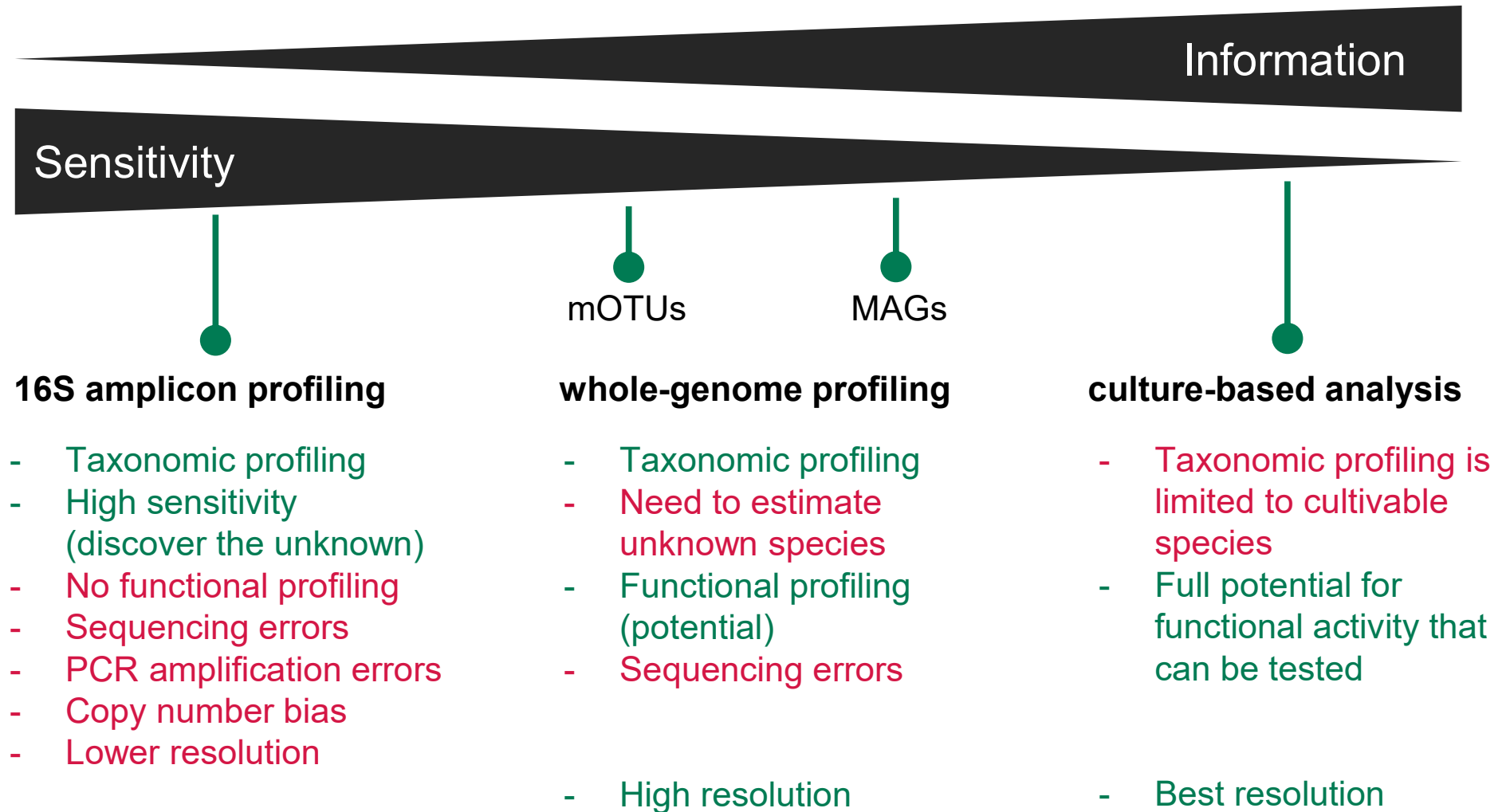


Functional profiling

- The other gene can be used to understand the functional potential



# Strengths and weaknesses of different approaches





# Take home message

- **Taxonomic profiling** is a fundamental step in the analysis of microbial communities
- **Incomplete reference databases** yield incomplete pictures (missing taxa) and tend to overestimate the abundances of known taxa
- 16S rRNA amplicon, shotgun sequencing and culture based approaches **are complementary**
- **16S rRNA amplicon** is a cheap solution that allow for sensitive taxonomic profiling
- **Shotgun sequencing** allows for a more precise taxonomic profiling (species and subspecies resolution) and functional profiling
- **Culture based approaches** are limited on the number of species that can study, but allows to fully investigate species properties
- **mOTUs** is one of the most competitive tools on the market for taxonomic profiling of metagenomes